



# The Certification of the Mass Fractions of As, Cd, Cu, Mn, Pb, Se and Zn in rice flour IRMM-804

A. Held



The mission of IRMM is to promote a common and reliable European measurement system in support of EU policies.

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Directorate-General Joint Research Centre  
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**A. Held**

## Summary

This report describes the preparation and certification of a rice flour Certified Reference Material (CRM) IRMM-804. The CRM was processed and certified by the European Commission, Directorate General Joint Research Centre, Institute for Reference Materials and Measurements (IRMM), Geel, Belgium.

The CRM was prepared from rice grown on Cd enriched water. After milling the resulting powder was filled in glass bottles containing 15 g of sample.

Certification of the CRM included testing of the homogeneity and stability of the material as well as the characterisation using an intercomparison approach.

The new CRM has been certified for its content of As, Cd, Cu, Mn, Pb and Zn, while the Se mass fraction is only given as "additional material information".

The main purpose of the material is to assess method performance, i.e. for checking accuracy of analytical results. As any reference material, it can also be used for control charts or validation studies.

	Mass Fraction	
	Certified value <sup>1)</sup> [mg/kg]	Uncertainty <sup>2)</sup> [mg/kg]
As	0.049	0.004
Cu	2.74	0.24
Mn	34.2	2.3
Pb	0.42	0.07
Cd	1.61	0.07
Zn	23.1	1.9
<p>1) Unweighted mean value of the means of accepted sets of data, each set being obtained in a different laboratory and/or with a different method of determination. The certified values are traceable to the SI.</p> <p>2) Expanded uncertainty with a coverage factor <math>k = 2</math> according to the Guide for the Expression of Uncertainty in Measurement, corresponding to a level of confidence of about 95 %.</p>		



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## Glossary

$\bar{y}$	average of all results of a homogeneity study
$\nu_{MS_{within}}$	degrees of freedom of $MS_{within}$
AAS	atomic absorption spectrometry
AFS	atomic fluorescence spectrometry
CRM	certified reference material
CVAFS	cold vapour AFS
ETAAS	electrothermal AAS
FAAS	flame AAS
GFAAS	graphite furnace AAS
HG	hydride generation
HGAAS	hydride generation AAS
HGAFS	hydride generation atomic fluorescence spectrometry
ICP-MS (Q / HR)	inductively coupled plasma mass spectrometry (quadrupole / high resolution i.e. sector field)
ICP-OES	inductively coupled plasma optical emission spectrometry
ID	isotope dilution technique
INAA	instrumental NAA
$k$	coverage factor
$k_0$ -NAA	NAA using the $k_0$ -method for quantification
$MS_{among}$	mean square among bottles from an ANOVA
$MS_{within}$	mean square within a bottle from an ANOVA
$n$	average number of replicates per bottle
NAA	neutron activation analysis
RNAA	radiochemical NAA
$s_{bb}$	between-unit variability
SI	Système International d'unités (international system of units)
$s_{meas}$	measurement variability
$s_{method}$	method variability
TIMS	thermal ionization mass spectrometry using the isotope dilution technique
$u_{bb}$	uncertainty related to a possible between-bottle inhomogeneity
$u_{bb}^*$	uncertainty of inhomogeneity that could be hidden by method repeatability
$u_c$	combined uncertainty of the certified value
$u_{c,bb}$	combined uncertainty of the between-unit measurement
$u_{char}$	uncertainty of the characterisation

$U_{CRM}$	expanded uncertainty of a certified value
$U_{ind}$	expanded uncertainty of an indicative value
$u_{Its}$	uncertainty of stability



## 1. Introduction

Food safety is a policy priority for the European Commission. This is reflected by a White Paper on Food Safety published in January 2000 [1]. It outlines a new food policy to be established in the years to come. This includes amongst others the setting up of a new food safety legislation covering the entire food chain from animal feed production to the final product on the consumers' plate. One measure was the Commission Regulation setting maximum levels for certain contaminants in foodstuffs [2], covering nitrate, mycotoxins, 3-monochloropropane-1,2-diol and heavy metals, Pb, Cd and Hg in particular. In addition, a Directive laying down sampling methods and methods of analysis [3] has been published. This directive requests the use of certified reference materials where possible.

Rice is staple food in large parts of the world, particularly Asia. Nevertheless, consumption as well as production have been growing significantly over the past decades in Europe. At present, roughly 2.6 million tons are produced in Europe, with a consumption of around 3.5 million tons [4].

Rice is one of the foodstuffs for which maximum levels for lead and cadmium have been set in the Commission Regulation [2]. Besides lead and cadmium, the elements copper, zinc, manganese, selenium and arsenic have been included in the certification of the rice flour reference material. Copper, zinc, manganese and selenium are essential trace elements, but also an excess of these elements can cause adverse effects. The European Commission has therefore requested the Scientific Committee for Food (SCF) to review the upper level on the daily intake for copper [5], zinc [6], manganese [7] and selenium [8]. Arsenic has been included in the certification as a potentially toxic element.

The rice flour was also used as test material in an interlaboratory comparison (IMEP-19 [9, 10]) and a Comité international des poids et mesures - Comité consultatif pour la quantité de matière – métrologie en chimie (CIPM - CCQM) key comparison CCQM-K24 [11] and pilot study CCQM-P29 [12] as well as an EUROMET comparison [13]. Results from the CCQM key comparison and pilot study were used in this certification with permission of the participants.

## 2. Participants

Participant	Activity <sup>1)</sup>
Bureau National de Métrologie et le Laboratoire National d'Essais (BNM – LNE), Paris, FR	C
Bundesanstalt für Materialforschung und –prüfung (BAM), Berlin, DE	C
Centro Nacional de Metrología (CENAM), Querétaro, MX	C
Central Science Laboratory (CSL), Sand Hutton, GB	C
Centre National de la Recherche Scientifique (CNRS), Service Central d'Analyse, Vernaison, FR	C
Centro de Energia Nuclear na Agricultura (CENA), São Paulo, BR	C
CSIR – National Metrology Laboratory, Pretoria, ZA	C
Eidg. Materialprüfungs- und Forschungsanstalt (EMPA), St. Gallen, CH	C
Institute Jožef Stefan, Department of Environmental Sciences, Ljubljana, SI	C
European Commission, DG JRC, Institute for Reference Materials and Measurements (EC-JRC-IRMM), Geel, BE	P, H, C
Korea Research Institute of Standards and Science (KRISS), KR	C
LGC Limited, Teddington, GB	C
National Analytic Reference Laboratory (NARL), Pymble, AU	C
National Metrology Institute of Japan (NMIJ), Tsukuba, JP	P, C
National Research Center for Certified Reference Materials (NRC CRM), Beijing, CN	C
National Research Council (NRC), INMS, Ottawa, CA	C
National Institute of Standards and Technology (NIST), Gaithersburg, US	C
Nederlands Meetinstituut (NMI) VSL, Delft, NL	C
Nuclear Research and consultancy Group (NRG) Petten, Petten, NL	C
Physikalisch-Technische Bundesanstalt (PTB), Braunschweig, DE	C
Rijksuniversiteit Gent, Gent, BE	C
Studiecentrum voor Kernenergie – Centre d'Etude de l'Energie Nucléaire (SCK-CEN), Mol, BE	S, H, C
Umweltbundesamt Wien, Wien, AT	C
Università di Pavia, Dipartimento di Chimica Generale, Nuclear Chemistry Section, Pavia, IT	C
University of Plymouth, Department of Environmental Sciences, Plymouth, GB	H, S, C
Vlaamse Instelling voor Technologisch Onderzoek (VITO), Mol, BE	C
WRc-NSF Ltd., Medmenham, GB	C

<sup>1)</sup>Activity: P: Processing of material, H: Homogeneity study, S: Stability study, C: Characterisation

### 3. Processing of the material

The rice flour used for IRMM-804 was provided by the National Metrology Institute of Japan (NMIJ) and originates from rice grown in cadmium contaminated water. The high lead content of this rice compared to published data [14] also suggests a contamination of the water or the soil with lead. The rice was milled and bottled in units of 60 g at NMIJ. At IRMM the rice flour was redistributed into units of 15 g and bottled in amber 60 mL glass bottles with a crimp seal. A total of 1526 bottles was produced.

The particle size distribution of the material was assessed using a particle size analyser with a Helos measuring device (Sympatec, Germany). A typical particle size distribution is shown in Figure 1. The top particle size was shown to be 500  $\mu\text{m}$ . The water content at the time of bottling was determined to be  $4.5 \pm 0.2 \%$  ( $n = 20$ ) by volumetric Karl Fisher titration. The water activity was determined to be  $0.102 \pm 0.005$  ( $n = 20$ ) using a water activity meter (Aqualab CX3, Decagon, USA). This low value indicates that the material is hygroscopic.

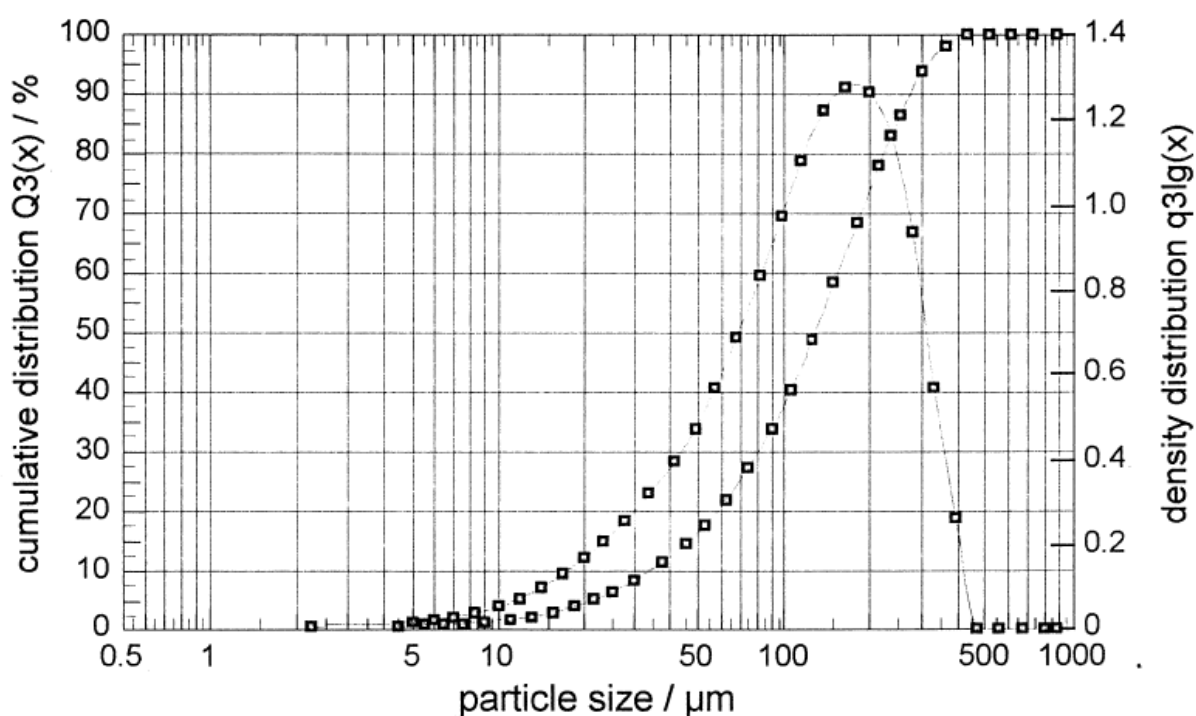


Figure 1: Typical particle size distribution of IRMM-804

## 4. Homogeneity

### 4.1. Between bottle homogeneity

The between bottle homogeneity is tested to ensure that the certified values of the CRM are valid for all bottles of the material, within the stated uncertainty. This was carried out using ICP-MS (As, Cd, Cu, Mn, Pb, Se, Zn), ICP-OES (Mn, Zn) and ETAAS (Cu, Cd, Pb) after pressure digestion with HNO<sub>3</sub> and HF.

For the elements Cd, Cu, Mn, Pb and Zn, 12 bottles were selected throughout the produced batch at approximately regular intervals, they were analysed for each analyte, three replicates have been measured per bottle. For As, the original measurements by ICP-MS were hampered by interferences. Instead, the k<sub>0</sub>-NAA data that was available for short- and long-term stability was also used to evaluate homogeneity. This is a valid approach, provided that no instability is detected (see Sections 5.1 and 5.2). This resulted in a dataset with measurements on 18 bottles which were analysed twice.

For Cd, Cu and Pb additional measurement data obtained by ID-ICP-MS was available. Using this method, 10 bottles have been measured, 5 bottles with 2 replicates per bottle, and 5 bottles with 3 replicates per bottle. This also allowed evaluation of the data as described below.

The results of these measurements were evaluated using a method described by Linsinger *et al.* [15] as described in the following paragraphs:

The obtained data is first tested whether they follow a normal, or at least unimodal distribution. This is done by visual inspection of normal probability plots and histograms. If the data does not follow at least a unimodal distribution, the calculation of standard deviations is doubtful or impossible.

The results were then evaluated by a one-way analysis of variance (ANOVA). From the results of the ANOVA calculation, the following figures are calculated:

Method repeatability ( $RSD_{method}$ ) expressed as a relative standard deviation is given as follows:

$$RSD_{method} = \frac{\sqrt{MS_{within}}}{\bar{y}}$$

$MS_{within}$ : mean square within a bottle from an ANOVA

$\bar{y}$ : average of all results of the homogeneity study

Between-bottle variation ( $s_{bb}$ ) expressed as a relative standard deviation is given by the following equation:

$$s_{bb} = \frac{\sqrt{\frac{MS_{among} - MS_{within}}{n}}}{\bar{y}}$$

$MS_{among}$ : mean square among bottles from an ANOVA

$n$ : average number of replicates per bottle

The inhomogeneity that can be hidden by method repeatability which is used as the minimum uncertainty contribution from homogeneity is defined as follows:

$$u_{bb}^* = \frac{RSD_{method}}{\sqrt{n}} \sqrt[4]{\frac{2}{v_{MS_{within}}}}$$

$\nu_{MS_{within}}$  : degrees of freedom of  $MS_{within}$

The larger value of  $s_{bb}$  or  $u_{bb}^*$  are used as uncertainty contribution for homogeneity,  $u_{bb}$ .

The results of the measurements are shown in Annex 2, Table 9 to Table 24. The statistical evaluation is summarised in Table 1. Where more than one result is available, the smaller value is used for calculating the uncertainty of the certified value (indicated by \* in the table) as in most cases the uncertainty contribution for homogeneity is determined by the method repeatability.

**Table 1: Results of the homogeneity study.  $u_{bb}$  as used for calculating the uncertainty of the certified values indicated with \*.**

element	method	$s_{bb}$ / %	$u_{bb}^*$ / %	$u_{bb}$ / %
As	ICP-MS	17	15	17
As	$k_0$ -NAA	0.97	0.93	0.97*
Cd	ETAAS	---	1.6	1.6
Cd	ICP-MS	---	1.5	1.5
Cd	ID-ICP-MS	0.30	0.16	0.30*
Cu	ETAAS	0.44	1.5	1.5
Cu	ICP-MS	---	1.5	1.5
Cu	ID-ICP-MS	0.83	0.66	0.83*
Mn	ICP-MS	---	1.6	1.6
Mn	ICP-OES	0.24	0.69	0.69*
Pb	ETAAS	2.4	1.9	2.4
Pb	ICP-MS	---	1.1	1.1*
Pb	ID-ICP-MS	1.1	0.48	1.1
Zn	ICP-MS	---	1.5	1.5
Zn	ICP-OES	---	0.68	0.68*

The data obtained for Pb is somewhat inconsistent, the data obtained by ETAAS results in an  $s_{bb}$  of 2.4 %, while with ICP-MS and ID-ICP-MS the data obtained results in  $s_{bb}$  and  $u_{bb}^*$  both of 1.1 %. Due to the good agreement of the ICP-MS and ID-ICP-MS data, a value of 1.1 % is used in the further evaluation.

#### 4.2. Micro-homogeneity, minimum sample intake

The minimum sample intake is the minimal mass of sample that shall to be used in an analysis, to ensure that the certified value is valid within its uncertainty. The micro-homogeneity of the materials was assessed using solid sampling ZAAS [16] for Cd, Cu, Pb and Zn. Sample intakes of about 0.3 – 0.6 mg were used for Cd, Cu and Pb, for Zn sample intakes of about 1.2 mg were used. 160 replicate measurements were carried out per element.

The data has been evaluated according to the following equation [17]:

$$M = \left( \frac{k'_2 \cdot s_m}{u_{target}} \right)^2 \cdot m$$

With  $M$  minimum sample mass,  $k'_2$  factor for the two-sided 95 % tolerance limits for a normal distribution,  $s_m$  relative standard deviation of the homogeneity experiment,  $u_{target}$  maximum relative uncertainty for sampling (this value should not exceed the uncertainty of the certified value) and  $m$  average mass used during the measurements.

The resulting minimum sample masses are summarised in Table 2.

**Table 2: Minimum sample masses for a target uncertainty of 5 % as determined by solid sampling AAS**

	$M$ (mg)	$s_m$	$m$ (mg)
Cd	22	0.135	0.63
Cu	15	0.146	0.36
Pb	19	0.160	0.39
Zn	12	0.069	1.24

For the other elements, the minimum sample intake was estimated from the characterisation study. Typical sample intakes used by the participating laboratories are used as minimum sample intake.

For As, Mn and Se sample intakes range from 0.2 to 1 g, most laboratories used 0.3 to 0.4 g.

For the four elements Cd, Cu, Pb and Zn a minimum sample intake of 20 mg can be considered acceptable, while for the elements As, Mn and Se the data of the characterisation study suggests that acceptable results can be obtained on samples of 200 mg or more.

There is no reason to assume that the elements As, Mn and Se are less homogeneously distributed than the elements Cd, Cu, Pb and Zn. Therefore, the overall minimum sample intake for this material, valid for all elements investigated, is set to 100 mg.

## 5. Stability studies

### 5.1. Short-term stability

The short-term stability is tested to establish dispatch conditions for the material. During transport, especially in summer time, quite high temperatures can be reached. This is simulated in the short-term stability study.

The short-term stability has been assessed using an isochronous design [18]. Samples have been stored at 18 and 40 °C for 0, 2, 4 and 8 weeks. The reference temperature was set to -20 °C. Due to problems with the As measurements, the short-term stability test was repeated for this element using a test temperature of 60 °C and a reference temperature of -20 °C.

The samples have been analysed by ICP-MS (As, Cd, Cu, Mn, Pb, Zn), ICP-OES (Mn, Zn) and ETAAS (Cu, Cd, Pb) after pressure digestion with HNO<sub>3</sub> and HF, under repeatability conditions. For As, the original measurements by ICP-MS were hampered by interferences, and were repeated. New measurements were carried out by k<sub>0</sub>-NAA on a new set of samples using 60°C as test temperature. The measurements for Se were not repeated, as the Se value is not certified and only given as additional material information.

The obtained data was first tested whether they follow a normal, or at least unimodal distribution. This is done by visual inspection of normal probability plots and histograms. If the data does not follow at least a unimodal distribution, the calculation of standard deviations is doubtful or impossible.

The data points obtained were plotted against storage time at the test temperature and the regression line calculated. The slope of the regression line was then tested for statistical significance. In Table 3 the results of these tests have been summarised. Only on two occasions this test indicated a significant slope. Nevertheless, in both cases the same samples have been measured additionally by another method, where no significant slope could be detected. The conclusion is therefore that the material is stable at 18°C and 40°C for up to 8 weeks.

**Table 3: Results of the short-term stability test**

Element	Method	18°C Significance	40°C Significance	60°C Significance
As	k <sub>0</sub> -NAA			no
Cd	ICP-MS	yes	no	
Cd	AAS	no	no	
Cu	ICP-MS	no	no	
Cu	GFAAS	no	no	
Mn	ICP-MS	no	no	
Mn	ICP-OES	no	no	
Pb	ICP-MS	no	yes	
Pb	GFAAS	no	no	
Zn	ICP-MS	no	no	
Zn	ICP-OES	no	no	

It can be concluded that the samples can be safely dispatched under conditions where the temperatures do not exceed 40 °C for up to 8 weeks, i.e. at ambient temperature.

## 5.2. Long-term stability

The long-term stability is tested to establish the shelf life of the CRM. The CRM is tested over a certain period of time, from which a prediction for the future is calculated. The samples have been analysed using ICP-MS (As, Cd, Cu, Mn, Pb, Se, Zn), ICP-OES (Mn, Zn) and ETAAS (Cu, Cd, Pb) after pressure digestion with HNO<sub>3</sub> and HF, under repeatability conditions. Again, for As additional measurements using k<sub>0</sub>-NAA were carried out. The Se data could not be used for the evaluation of the long-term stability. Since no certified value is established for this element, no additional measurements were carried out.

Also the long-term stability has been assessed using an isochronous design [18]. In this case samples have been stored at 4 °C for up to 24 months. The reference temperature was set to –20 °C. Also here measurements for As had to be repeated. Since the long-term stability could not easily be repeated due to the long time periods involved, a different approach was chosen. In parallel to the isochronous stability study at 4 °C, a second study at 18 °C had been conducted, but those samples had not been measured. This set of samples was now measured by k<sub>0</sub>-NAA for As. Additionally, two bottles of the material stored at the regular storage temperature (4 °C) were measured in comparison to two bottles of the material stored at the reference temperature for 46 months. This could be considered as a two-point isochronous stability study covering 46 months.

The data points were plotted against time and the regression line calculated. In all cases the slope of the regression line was found to be insignificant. The uncertainty of stability  $u_{lts}$  of the materials is then calculated for the required shelf life as:

$$u_{lts} = \frac{RSD}{\sqrt{\sum (x_i - \bar{x})^2}} \cdot x$$

with  $RSD$  being the relative standard deviation of all results of the stability study,  $x_i$  being the time point for each replicate,  $\bar{x}$  being the average of all time points and  $x$  being the proposed shelf life (48 months at 4 °C in this case).



The results are summarised in Table 4.

**Table 4: Uncertainty contribution of the long-term stability,  $u_{lts}$ , for a shelf life of 48 months. Values included in the final evaluation of the combined uncertainty of the certified values are indicated by \*.**

Element	Method	$u_{lts}$ (%)	comment
As	ICP-MS	31.8	
As	$k_0$ -NAA	4.2	Isochronous study at 18 °C, 24 months
As	$k_0$ -NAA	2.3*	Isochronous study at 4 °C, 46 months
Cd	GFAAS	3.6	
Cd	ICP-MS	1.9*	
Cu	GFAAS	3.1*	
Cu	ICP-MS	6.1	
Pb	GFAAS	10.5	
Pb	ICP-MS	6.4*	
Zn	ICP-OES	4*	
Zn	ICP-MS	5.7	
Mn	ICP-OES	1.9*	
Mn	ICP-MS	2.9	

All slopes of the regression lines that were calculated for the different analytes were not significantly different from zero. Therefore a  $u_{lts}$  could be established for all analytes for 48 months at 4 °C. The material will be subjected to IRMM's regular stability monitoring programme to ensure stability beyond the initial 48 months.

## **6. Characterisation**

### **6.1. Approach**

The certification of the material has been carried out using a laboratory intercomparison approach. In that intercomparison different laboratories used different methods to determine the analyte content in the rice flour. Nevertheless, dry mass determination had to be carried out according to a described procedure.

For the characterisation of the elements Cd and Zn results of the CCQM key comparison K24 [11] and the pilot study P29 [12] have been used. In these comparisons the same material (just differently labelled) has been used. One laboratory could only participate to the pilot study or the key comparison, thus no duplication of results could occur. Permission to use the data have been obtained from all but one participant of the key comparison and pilot study. The data of that one participant were consequently not used.

For the other analytes (As, Cu, Mn, Pb and Se) a group of laboratories has been recruited to carry out the measurements.

### **6.2. Dry mass determination**

For all measurements carried out for the certification of IRMM-804, the following protocol for dry mass determination was prescribed for the participants:

Dry mass determination should be carried out on separate subsamples, one subsample per independent digestion. Weighing of the samples for dry mass determination and the analysis must be done at the same time to avoid differences in moisture due to the hygroscopicity of the rice flour. Dry mass determination should be carried out by drying in a ventilated oven at 85 °C for at least 12 hours, until constant weight is reached. Constant weight is reached when two consecutive net sample weights differ by less than 0.5 %.

The protocol followed by the CCQM participants required a minimum sample mass of 1 g and defined constant weight when two consecutive net sample weights differed by less than 0.001 g [11, 12]. The IRMM protocol for IRMM-804 is more general than the CCQM protocol, but they agree. Therefore all sets of data are comparable with respect to the dry mass determination.

### **6.3. Methods used**

The methods used in the characterisation study are summarised in Annex 1. Each participant received 2 bottles of the material, and was requested to provide 6 independent results, three per bottle. In addition, the water content had to be determined on the same number of separate subsamples.

### **6.4. Evaluation of results**

Individual results of the participants, grouped per element are displayed in Annex 4. The obtained data was subjected to outlier tests (Dixon, Nalimov, Grubbs, Cochran). Based on these tests and a visual inspection of the data some outliers could be identified. Additionally the analysis reports from the participants were checked for compliance with the requested

analysis. No data was excluded from the evaluation based on outlier tests alone. Only if a sound technical reason was given, datasets were excluded.

To combine the data from the different labs, the certified value is calculated as the mean of the laboratory means. The contribution of the characterisation to the uncertainty of the certified value  $u_{\text{char}}$  is estimated as the standard deviation of the mean of the laboratory means. The resulting data is summarised in Table 5.

**Table 5: Summary of data of the characterisation**

Analyte	Number of independent, valid datasets	Mean of laboratory means (mg / kg)	$u_{\text{char}}$ (%)
As	8	0.0494	2.8
Cu	9	2.741	2.8
Mn	9	34.17	2.6
Pb	9	0.418	3.3
Se	5	0.038	9.7
Cd	17	1.615	0.4
Zn	13	23.06	0.5

In Table 6 some data sets that were found to be outliers or non-compliant are listed, together with the action taken.

**Table 6: Datasets that were found to be outliers or showed other non-compliances**

Element	Lab	Description of problem	Action taken
All	C ICP-MS and C F-AAS and C HG-AAS	The lab used the same digests for all methods, thus violating the requirement of independence	Data from all methods were used for calculating the average value per element, nevertheless, they are only counted as one independent method for the calculation of $U_{char}$ .
All	J INAA	Lab was identified as outlier for 3 out of 4 elements measured and significantly deviates from the results of other labs for these 3 elements	As the findings of lab J INAA are not supported by the results of lab F INAA, the outlying data is not a method specific problem. As it seems to be a problem of the laboratory, all data of lab J INAA are excluded from evaluation.
As	H ICP-MS	As concentration was below LOQ of that lab	Value is excluded
Cu	H GFAAS	Was identified as outlier by all tests, lab reported possible matrix effect	Value is excluded
Pb	I ICP-MS	Was identified as outlier using the Nalimov t-test at 95 % confidence level	Review of the report of that lab and the measurement method did not point to any problems, value is retained.
Se	D ICP-MS and I ICP-MS	Reported 'less than' values	'Less than' values are excluded
	H ICP-MS	Se concentration below LOQ of that lab	Value is excluded
Cd	CCQM 7 ID ICP-MS	Was identified as outlier by all tests	Lab also reported large uncertainty of their measurement that overlaps with the certified uncertainty. Value is retained.
Zn	CCQM 7 ID ICP-MS	Was identified as outlier by all tests	Lab also reported large uncertainty of their measurement that overlaps with the certified uncertainty. Value is retained.

## 7. Certified / indicative values

### 7.1. Certified values and uncertainties

The certified values result from the characterisation study (Table 5). The uncertainty of the certified values contains contributions of the characterisation  $u_{char}$ , the homogeneity  $u_{bb}$  and the long-term stability  $u_{lts}$ .

The different contributions to the CRM uncertainty are combined using equation (1).

$$U_{CRM} = k \cdot \sqrt{u_{char}^2 + u_{bb}^2 + u_{lts}^2} \quad (1)$$

The expanded uncertainty of the certified value  $U_{CRM}$  is calculated with a coverage factor of  $k = 2$ , representing a level of confidence of approximately 95%.

**Table 7: Summary of the certified values and uncertainties**

	Certified value (mg/kg)	$U_{CRM}$ k = 2 (mg/kg)	$U_{CRM}$ (%)	$u_{char}$ (%)	$u_{bb}$ (%)	$u_{lts}$ (%)
As	0.049	0.004	8	2.8	1.0	2.3
Cu	2.74	0.24	9	2.8	0.8	3.1
Mn	34.2	2.3	7	2.6	0.7	1.9
Pb	0.42	0.07	17	3.3	1.1	6.4
Cd	1.61	0.07	4	0.4	0.3	1.9
Zn	23.1	1.9	8	0.5	0.7	4.0

### 7.2. Additional material information

The Se data obtained from the participants of the characterisation does not show a good agreement between labs. Also only 5 independent datasets are available. Additionally, the data that was obtained in the homogeneity and stability study for Se was not good enough to be evaluated, although it can be assumed that Se should exhibit the same stability and homogeneity as the other elements investigated. Nevertheless, this prevents the use of these data to provide a certified value for Se. It is given as additional material information.

**Table 8: Additional material information**

	Value (mg/kg)	Number of labs
Se	0.038	5

This value is only reported as "additional material information" and should not be used for any quality control purposes.

### 7.3. Metrological traceability

Traceability of the certified values to the SI is ensured through the set-up of the characterisation. The participating laboratories used a number of different methods for the

sample preparation as well as for the final determination, thus eliminating any possibility of method dependent results. In addition, different calibrants have been used, including commercial standard solutions, CRMs and in-house gravimetrically prepared calibrants. Most laboratories also used matrix CRMs for quality control. In addition, the measurements for the characterisation of Cd and Zn have been carried out by national metrology institutes or designated laboratories, thus at a very high metrological level.

#### **7.4. Commutability**

Commutable CRMs must exhibit the same analytical behaviour for given methods as a real laboratory sample. The laboratories participating in the characterisation study have been selected such as to provide a large variety of analytical methods, regarding digestion, calibration and detection. The good agreement between the results obtained shows the commutability of the material. Nevertheless it has to be kept in mind that the certified reference materials might show a behaviour different from real samples, in particular during digestion, due to their small particle sizes in contrast to the possibly larger particle sizes encountered for real laboratory samples, and the intensive processing that these materials have undergone.

## 8. Instructions for use

The main purpose of the material is to assess method performance, i.e. for checking accuracy of analytical results. As any reference material, it can also be used for control charts or validation studies.

### Storage of the material

Samples should be stored refrigerated at 4 °C. Care should be taken to avoid moisture pickup once the bottles are open, as the material is hygroscopic. However, the European Commission cannot be held responsible for changes that happen during storage of the material at the customer's premises, especially of opened samples.

### Use of the material

The bottles should be shaken by turning upside down for at least two minutes before opening to ensure re-homogenisation of the content.

### Dry mass correction

Dry mass determination should be carried out on separate subsamples. Weighing of the samples for dry mass determination and the analysis must be done at the same time to avoid differences in moisture due to the hygroscopicity of the rice flour. Dry mass determination should be carried out by drying in a ventilated oven at 85 °C for at least 12 hours, until constant weight is reached. At this stage two consecutive net sample weights should differ by less than 0.5 %.

### Comparing an analytical result with the certified value

A result is unbiased if the combined uncertainty of measurement and certified value covers the difference between the certified value and the measurement result. To this end, the following steps are necessary:

- 1) Assessment of the measurement uncertainty: This uncertainty will depend whether accuracy of one individual result or accuracy of a method in general shall be assessed. Measurement uncertainty can be estimated from reproducibility data obtained during method validation. These reproducibility data do not comprise uncertainty of the calibration. This uncertainty has to be added. The uncertainty is then estimated as

$$u_{meas} = c \sqrt{u_R^2 + u_{st}^2}$$

$u_{meas}$  standard measurement uncertainty

$c$  concentration for which the uncertainty should be evaluated

$u_R$  relative uncertainty due to reproducibility (as taken from the validation study)

$u_{st}$  relative uncertainty of the calibration standard (mainly purity).

$u_{st}$  can be ignored if it is  $< 1/3 u_R$

- 2) Take the standard uncertainty of the certified value of the material in question ( $u_{CRM}$ ).  
E.g. for As in IRMM-804, this would be 0.002 mg/kg.

3) Combine the two uncertainties as  $u_c = \sqrt{u_{meas}^2 + u_{CRM}^2}$

4) The measurement result is not significantly biased if the difference between the measured result and the certified value is smaller than  $2 u_c$ .

#### Use in quality control charts

The material can be used for quality control charts. Different CRM-units will give the same result as heterogeneity was found negligible.

#### Use as a calibrant

It is not recommended to use matrix materials such as IRMM-804 as calibrants. If used as calibrant, the uncertainty of the certified value shall be taken into consideration in the final estimation of measurement uncertainty.



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## **Annexes**

### Annex 1: Summary of methods used

lab no	Method acronym	Sample mass (g)	Sample preparation	Calibrants	Instrumentation and measurement method	Elements
A	ID-ICP-MS	0.4	Microwave digestion using 0.3 g HF and 4.5 g HNO <sub>3</sub> , 20 min at 250 W	Spike solutions ( <sup>65</sup> Cu and <sup>206</sup> Pb) calibrated against gravimetrically prepared solutions of high purity Cu and Pb metal	ICP-MS (HR) operated at medium resolution (R = 3000), <sup>65</sup> Cu/ <sup>63</sup> Cu and <sup>208</sup> Pb/ <sup>206</sup> Pb ratios measured	Cu, Pb
B	ID-ICP-MS	0.5	Digestion in high pressure asher, using 2 mL HNO <sub>3</sub> and 2 mL H <sub>2</sub> O <sub>2</sub> , 90 min at 240 °C, 100 bar	Spike solution ( <sup>206</sup> Pb) Certipur	ICP-MS (Q), <sup>206</sup> Pb/ <sup>208</sup> Pb ratio measured, in addition <sup>204</sup> Pb/ <sup>208</sup> Pb, <sup>207</sup> Pb/ <sup>208</sup> Pb ratios measured in unspiked sample	Pb
C	HGAAS	1	Digestion using HNO <sub>3</sub>	Certified standards from a variety of manufacturers	HGAAS, external calibration	As, Se
	ICP-MS				ICP-MS (Q), external calibration with internal standard (Rh)	As, Se, Pb, Mn, Cu
	FAAS				FAAS	Pb, Mn, Cu
D	ICP-MS	0.4	Closed vessel microwave digestion using 5 mL HNO <sub>3</sub> , 10 min 700 to 1000 W, 10 min at 1000 W, after addition of 2 mL HF another 20 min at 1000 W	Merck ICP standard solutions	ICP-MS (Q), matrix matched external standard with internal standard, isotopes monitored: <sup>75</sup> As, <sup>113</sup> Cd, <sup>65</sup> Cu, <sup>55</sup> Mn, <sup>66</sup> Zn, <sup>82</sup> Se, <sup>208</sup> Pb	Pb, Cd, Se, As, Mn, Cu, Zn
E	k <sub>0</sub> -NAA	1	not applicable	IRMM-530	7 h irradiation, thermal neutron flux of 1.3·10 <sup>12</sup> cm <sup>-2</sup> s <sup>-1</sup> and 1.6·10 <sup>12</sup> cm <sup>-2</sup> s <sup>-1</sup> , Ge-detector	Mn, As, Cu
F	INAA	0.2 - 0.4	not applicable	Gravimetrically prepared solutions of high purity As and Se were dried into quartz ampoules	1 day irradiation for As, 14 days for Se, thermal neutron flux 6.6E12 cm <sup>-2</sup> s <sup>-1</sup> , Ge detector	Se, As

lab no	Method acronym	Sample mass (g)	Sample preparation	Calibrants	Instrumentation and measurement method	Elements
G	ICP-MS	0.3 -0.5	Open digestion using HClO <sub>4</sub> and HNO <sub>3</sub> for 20 h at 250 °C	SPEX certified solutions	ICP-MS (Q), external calibration with internal standards, isotopes monitored: <sup>75</sup> As, <sup>63</sup> Cu, <sup>65</sup> Cu, <sup>206</sup> Pb, <sup>207</sup> Pb, <sup>208</sup> Pb, <sup>82</sup> Se	Cu, Pb, Se
			Open digestion using H <sub>2</sub> SO <sub>4</sub> and HNO <sub>3</sub> for 20 h at 250 °C			As
	ICP-OES		Open digestion using HClO <sub>4</sub> and HNO <sub>3</sub> for 20 h at 250 °C		ICP-AES, external calibration, Mn: 257.611 nm, background correction	Mn
H	ICP-OES	0.5	Digestion under reflux using 5 mL HNO <sub>3</sub> , heat to 65 °C for 8 h, then to 95 °C for 8 h, addition of 0.1 mL HF, further heating to 95 °C for 8 h. Evaporation to near dryness and redissolution in HNO <sub>3</sub>	Certified single element calibration solutions (PrimAg-xtra, UK), NIES 10 CRM	Mn: 257.610 nm, Zn: 213.856 nm	Mn, Zn
	ETAAS				Cd: 283.3 nm, modifier 50 µg PO <sub>4</sub> + 3 µg Mg(NO <sub>3</sub> ) <sub>2</sub> , Cu: 324.8 nm, modifier 5 µg Pd + 3 µg Mg(NO <sub>3</sub> ) <sub>2</sub> , Pb 228.8 nm, modifier 50 µg PO <sub>4</sub> + 3 µg Mg(NO <sub>3</sub> ) <sub>2</sub>	Cd, Cu, Pb
	ICP-MS				ICP-MS (HR) (resolution 400: Cd, Pb, resolution 4000: Mn, Cu, Zn, resolution 10000: As, Se), external calibration with Rh internal standard, isotopes monitored: <sup>111</sup> Cd, <sup>208</sup> Pb, <sup>55</sup> Mn, <sup>63</sup> Cu, <sup>66</sup> Zn, <sup>75</sup> As, <sup>80</sup> Se	As, Cd, Cu, Mn, Pb, Se, Zn
I	ICP-MS	0.5	Closed vessel microwave digestion using 5 mL HNO <sub>3</sub>	Spectrosol and Aristar single element calibration standards and CRMs (NIST, BCR, NRCC)	ICP-MS (Q), external calibration with internal standard (Rh)	As, Se, Pb, Mn, Cu
J	INAA	0.2 - 0.5	not applicable	Johnson Matthey Specpure As <sub>2</sub> O <sub>3</sub> , MnO <sub>2</sub> , SeO <sub>2</sub> , Cu and Zn metal	INAA, irradiation As, Se, Zn: 10 h, Mn: 600 s, Cu: 100 s, neutron flux: 4·10 <sup>12</sup> cm <sup>-2</sup> s <sup>-1</sup> , intrinsic Ge detector	As, Cu, Mn, Se, Zn

lab no	Method acronym	Sample mass (g)	Sample preparation	Calibrants	Instrumentation and measurement method	Elements
K	ETAAS	0.2	Closed vessel microwave digestion using 4 mL HNO <sub>3</sub> , heat to 180 °C during 20 min, hold for 20 min, then cool down	SRM 3144 (NIST)	ETAAS using Zeeman background correction, Cu: 324.8 nm	Cu
	FAAS			SRM 3132 (NIST)	FAAS using a nitrous-oxide flame, Mn: 279.5 nm, background correction using a deuterium lamp	Mn
	HGAFS	0.2 – 0.25	Digestion at ambient pressure using 0.5 mL H <sub>2</sub> SO <sub>4</sub> and 1.5 mL of HNO <sub>3</sub> heated to 60 °C over night, then to 130 °C for 1 h. Afterwards addition of twice 2 mL H <sub>2</sub> O <sub>2</sub> and heating to 115 °C, addition of 0.1 mL V <sub>2</sub> O <sub>5</sub> in H <sub>2</sub> SO <sub>4</sub> and heating to 115°C, then reduction of Se(VI) to Se(IV) with 2.5 mL conc. HCl at 100 °C for 10 min	Prepared from Na <sub>2</sub> SeO <sub>3</sub> (Sigma) in the same media as samples	HGAFS	Se
	RNAA	0.4	As: Wet digestion of irradiated sample with carrier, using H <sub>2</sub> SO <sub>4</sub> , HNO <sub>3</sub> and H <sub>2</sub> O <sub>2</sub> . Addition of KI and extraction of AsI <sub>3</sub> into toluene and re-extraction into H <sub>2</sub> O  Se: Destruction of the irradiated sample using Mg(NO <sub>3</sub> ) <sub>2</sub> , reduction with 6 M HCl, derivatization with 4-nitro-1,2-diaminobenzene and extraction of Se chelate in toluene	Merck single elements standard solutions	Well type and coaxial HPGe detectors, neutron flux 1.1·10 <sup>12</sup> cm <sup>-2</sup> ·s <sup>-1</sup>  As: 20 h irradiation, measured at 559.1 keV  Se: 40 h irradiation, measured at 400.7 keV	As, Se

lab no	Method acronym	Sample mass (g)	Sample preparation	Calibrants	Instrumentation and measurement method	Elements
L	k <sub>0</sub> -NAA	0.6	not applicable	IRMM-530, Alfa Aesar single element standard solutions	7 h irradiation, thermal neutron flux of $3 \cdot 10^{11} \text{ cm}^{-2} \text{ s}^{-1}$ (As), 15 min irradiation, thermal neutron flux of $2 \cdot 10^{11} \text{ cm}^{-2} \text{ s}^{-1}$ (Mn),	Mn, As
M	ID-ICP-MS	0.2	Microwave digestion using 4.5 mL HNO <sub>3</sub> and 0.5 mL HF, microwave heating for 40 min using various power settings	IRMM-632 <sup>65</sup> Cu spike solution, IRMM-633 natural Cu solution	ICP-MS (Q), <sup>63</sup> Cu/ <sup>65</sup> Cu ratio measured	Cu
CCQM-						
1	INAA	0.25	not applicable	Gravimetrically prepared solutions of high purity metals, SRM 3108	16 h irradiation, neutron flux $3.1 \cdot 10^{13} \text{ cm}^{-2} \text{ s}^{-1}$	Cd, Zn
2	k <sub>0</sub> -NAA	0.56	not applicable	Gravimetrically prepared solutions of Cd and Zn metal, k <sub>0</sub> -method	Thermal neutron flux $1.2 \cdot 10^{13} \text{ cm}^{-2} \text{ s}^{-1}$	Cd, Zn
3	ID-ICP-MS	0.5	Closed vessel microwave digestion, 6 mL H <sub>2</sub> O <sub>2</sub> , 10 mL HNO <sub>3</sub> , stepwise heating to 190 °C during 40 min, keep at 190 °C for 30 min, evaporate to dryness, add 6 mL H <sub>2</sub> O <sub>2</sub> , 10 mL HNO <sub>3</sub> , heat to 210 °C within 30 min, keep for 20 min (microwave)	Spike solutions calibrated with SRM 746 and SRM 682	ICP-MS (HR), double focussing magnetic sector ICP-MS, single collector (electron multiplier), <sup>111</sup> Cd/ <sup>114</sup> Cd and <sup>67</sup> Zn/ <sup>64</sup> Zn ratio measured	Cd, Zn
4	ID-ICP-MS	0.2 - 0.25	Closed vessel microwave digestion, 4 mL HNO <sub>3</sub> , 1 mL H <sub>2</sub> O <sub>2</sub>		ICP-MS (Q), <sup>111</sup> Cd/ <sup>106</sup> Cd and <sup>68</sup> Zn/ <sup>67</sup> Zn ratio measured	Cd, Zn

lab no	Method acronym	Sample mass (g)	Sample preparation	Calibrants	Instrumentation and measurement method	Elements
5	ID-ICP-MS	0.5	Closed vessel microwave digestion, 5 mL HNO <sub>3</sub> , 0.5 mL HCl, heating for 35 min with max. pressure of 35 bar	Spike solutions ( <sup>106</sup> Cd and <sup>67</sup> Zn) were calibrated against a gravimetrically prepared solution of high purity Cd and Zn, and against SRM 3108 and SRM 3168a; NIES10c and SRM 1568a as matrix CRMs	ICP-MS (HR), double focussing magnetic sector ICP-MS in low resolution, isotopes monitored: <sup>111</sup> Cd, <sup>106</sup> Cd, <sup>67</sup> Zn and <sup>66</sup> Zn	Cd, Zn
6	ID-ICP-MS	0.25	Closed vessel microwave digestion, 5 mL HNO <sub>3</sub> , 1 mL HF and 0.2 mL H <sub>2</sub> O <sub>2</sub> , heating at different power settings for 90 min	Spike solutions ( <sup>111</sup> Cd and <sup>67</sup> Zn), calibrated by reverse IDMS using a primary assay standard	ICP-MS (Q), <sup>114</sup> Cd/ <sup>111</sup> Cd and <sup>66</sup> Zn/ <sup>67</sup> Zn ratio measured	Cd, Zn
7	ID-ICP-MS	0.45	Microwave digestion using 10 g HNO <sub>3</sub>		ICP-MS (HR)	Cd, Zn
8	ID-ICP-MS	0.25	Microwave digestion using 4.5 mL HNO <sub>3</sub> and 0.5 g HF	Spike solution ( <sup>111</sup> Cd) was calibrated against a solution of high purity Cd metal	ICP-MS (HR)	Cd
9	ID-ICP-MS	0.4	Microwave digestion using 1 mL H <sub>2</sub> O and 3 mL HNO <sub>3</sub> , stepwise heating for 30 min with power up to 600 W, addition of 0.5 mL HClO <sub>4</sub> , 0.5 mL HF, stepwise heating for 30 min with power up to 600W; for Zn matrix separation using an ion exchange column	Spike calibration ( <sup>111</sup> Cd, <sup>68</sup> Zn) by reverse IDMS	ICP-MS (HR), isotopes monitored: <sup>111</sup> Cd, <sup>112</sup> Cd, <sup>66</sup> Zn, <sup>68</sup> Zn	Cd, Zn
10	ID-ICP-MS	0.25	Microwave digestion using 4.5 g HNO <sub>3</sub> , 0.5 g HF		ICP-MS (Q), <sup>110</sup> Cd/ <sup>111</sup> Cd ratio measured	Cd



lab no	Method acronym	Sample mass (g)	Sample preparation	Calibrants	Instrumentation and measurement method	Elements
10	ko-NAA	0.35	not applicable	IRMM-530	7 h irradiation, thermal neutron flux of $3 \cdot 10^{11} \text{ cm}^{-2} \text{ s}^{-1}$	Zn
11	ID-TIMS	0.4	Microwave digestion using 10 mL $\text{HNO}_3$ , 180 °C for 30 min, evaporate to dryness after addition of 0.5 mL $\text{HClO}_4$ ; matrix separation on an anion exchange column		TIMS (multicollector instrument), single Re filament, silica gel, $^{112}\text{Cd}/^{113}\text{Cd}$ and $^{67}\text{Zn}/^{66}\text{Zn}$ ratio measured	Cd, Zn
12	ID-ICP-MS	0.8	Microwave digestion using 5 mL $\text{HNO}_3$ , heating to 230 °C, max. 75 bar for 30 min	Spike: IRMM-621, Oak Ridge Zn66, calibrated against SRM 3108 and SRM 3168a	ICP-MS (HR), used in low resolution, $^{114}\text{Cd}/^{111}\text{Cd}$ measured; ICP-MS (Q) with collision cell, $^{64}\text{Zn}$ and $^{66}\text{Zn}$ measured	Cd, Zn
13	ID-TIMS	0.3	Digestion using $\text{HNO}_3$ , matrix separation using an ion exchange resin		TIMS, $^{112}\text{Cd}/^{111}\text{Cd}$ ratio measured	Cd
14	ID-ICP-MS	0.5	Microwave digestion using 4 mL $\text{HNO}_3$ and 0.1 mL $\text{HClO}_4$ , after addition of 0.4 mL $\text{HClO}_4$ and 0.2 mL second microwave digestion	$^{111}\text{Cd}$ and $^{66}\text{Zn}$ used as spike	ICP-MS with collision cell, $^{112}\text{Cd}/^{111}\text{Cd}$ and $^{68}\text{Zn}/^{66}\text{Zn}$ ratio measured	Cd, Zn
15	ID-ICP-MS	0.5	Microwave digestion using $\text{HNO}_3:\text{H}_2\text{O}:\text{H}_2\text{O}_2$ (4:5:1)	$^{111}\text{Cd}$ used as spike	ICP-MS (HR), $^{112}\text{Cd}/^{111}\text{Cd}$ ratio measured	Cd
16	ID-ICP-MS	0.5 - 1	Closed vessel digestion using $\text{HNO}_3$ at 85 °C, extraction of Cd as iodide complex	$^{111}\text{Cd}$ spike (IRMM-621)	ICP-MS (MC)	Cd

## Annex 2: Data from homogeneity studies

**Table 9: Results for Cu by GFAAS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	3.66	3.46	3.33
179	3.52	3.59	3.35
282	3.60	3.54	3.81
486	3.53	3.48	3.37
687	3.65	3.11	3.79
791	3.65	3.38	3.82
894	3.50	3.24	3.40
994	3.53	3.22	3.30
1200	3.43	3.46	3.34
1302	3.57	3.43	3.35
1403	3.54	3.28	3.35
1507	3.34	3.42	3.31

**Table 10: Results for Cd by GFAAS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	1.55	1.59	1.63
179	1.50	1.59	1.64
282	1.52	1.77	1.42
486	1.53	1.59	1.52
687	1.56	1.47	1.56
791	1.59	1.50	1.59
894	1.58	1.56	1.52
994	1.61	1.65	1.58
1200	1.68	1.54	1.59
1302	1.63	1.63	1.61
1403	1.61	1.63	1.58
1507	1.77	1.73	1.49

**Table 11: Results for Pb by GFAAS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	0.461	0.504	0.462
179	0.477	0.534	0.454
282	0.467	0.496	0.443
486	0.438	0.503	0.431
687	0.454	0.497	0.456
791	0.457	0.505	0.452
894	0.528	0.516	0.549
994	0.460	0.462	0.495
1200	0.462	0.510	0.435
1302	0.459	0.494	0.409
1403	0.510	0.519	0.464
1507	0.482	0.502	0.490

**Table 12: Results for As by ICP-MS (results are expressed in mg/kg; n.d.: not detected, concentration below the limit of detection of the method)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	0.0153	n.d.	n.d.
179	0.0169	n.d.	n.d.
282	0.0198	n.d.	n.d.
486	0.0121	0.0102	n.d.
687	0.0286	n.d.	0.0271
791	0.0166	0.0101	0.0138
894	n.d.	0.0229	0.0145
994	0.0234	0.0120	0.0315
1200	0.0199	0.0165	0.0212
1302	0.0378	n.d.	0.0181
1403	0.0125	0.0181	n.d.
1507	n.d.	0.0151	0.0126

**Table 13: Results for Cd by ICP-MS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	1.66	1.57	1.51
179	1.56	1.61	1.34
282	1.63	1.54	1.54
486	1.61	1.56	1.34
687	1.51	1.53	1.57
791	1.56	1.57	1.58
894	1.50	1.41	1.54
994	1.54	1.66	1.57
1200	1.57	1.57	1.43
1302	1.60	1.58	1.56
1403	1.52	1.53	1.50
1507	1.44	1.55	1.54

**Table 14: Results for Cu by ICP-MS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	2.49	2.44	2.32
179	2.40	2.47	2.02
282	2.48	2.38	2.43
486	2.45	2.48	2.26
687	2.52	2.36	2.46
791	2.45	2.47	2.46
894	2.42	2.21	2.54
994	2.47	2.61	2.49
1200	2.51	2.48	2.17
1302	2.58	2.47	2.54
1403	2.41	2.40	2.45
1507	2.27	2.48	2.41

**Table 15: Results for Mn by ICP-MS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	37.1	37.2	33.6
179	35.0	36.7	30.6
282	36.8	34.3	35.2
486	36.5	35.2	30.6
687	36.7	34.9	36.2
791	35.5	36.6	35.3
894	35.6	32.2	37.1
994	36.2	38.1	36.9
1200	36.2	36.4	32.6
1302	37.6	37.1	36.4
1403	37.0	35.4	37.2
1507	33.8	36.1	36.5

**Table 16: Results for Pb by ICP-MS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	0.464	0.458	0.420
179	0.442	0.456	0.441
282	0.444	0.428	0.450
486	0.446	0.448	0.445
687	0.452	0.428	0.449
791	0.443	0.449	0.445
894	0.432	0.492	0.447
994	0.439	0.465	0.443
1200	0.439	0.435	0.406
1302	0.458	0.441	0.450
1403	0.441	0.454	0.459
1507	0.412	0.441	0.446

**Table 17: Results for Se by ICP-MS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	0.122	0.077	0.112
179	0.094	0.119	0.101
282	0.091	0.110	0.133
486	0.164	0.119	0.106
687	0.128	0.123	0.115
791	0.144	0.098	0.164
894	0.137	0.110	0.106
994	0.156	0.132	0.138
1200	0.110	0.077	0.153
1302	0.105	0.130	0.133
1403	0.114	0.112	0.132
1507	0.114	0.125	0.088

**Table 18: Results for Zn by ICP-MS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	21.4	21.2	19.8
179	20.8	21.8	18.1
282	21.7	20.0	20.8
486	21.3	21.4	19.3
687	21.2	20.3	21.5
791	21.2	21.5	21.1
894	20.2	19.3	21.4
994	20.7	23.2	21.8
1200	21.5	21.3	19.0
1302	22.3	21.6	22.1
1403	20.1	20.4	20.7
1507	19.5	21.7	20.4

**Table 19: Results for Mn by ICP-OES (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	37.2	34.0	34.6
179	35.5	35.7	35.3
282	34.9	34.2	35.3
486	33.2	34.9	34.2
687	35.3	34.1	35.2
791	35.7	35.4	34.6
894	34.6	35.9	35.6
994	34.5	33.9	35.6
1200	35.5	35.4	35.6
1302	36.3	35.4	34.9
1403	34.8	34.9	35.3
1507	34.3	35.3	33.8

**Table 20: Results for Zn by ICP-OES (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
77	24.4	22.4	22.7
179	22.8	22.3	22.7
282	23.5	22.5	22.9
486	22.3	22.6	23.1
687	23.3	22.9	23.3
791	22.8	22.7	22.9
894	23.2	23.5	23.3
994	22.8	22.9	22.7
1200	23.3	22.8	23.8
1302	23.1	22.6	23.4
1403	23.0	22.8	22.4
1507	22.9	24.4	23.2

**Table 21: Results for Pb by ID-ICP-MS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
18	0.413	0.415	
77	0.435	0.424	
156	0.419	0.418	
164	0.403	0.411	
187	0.427	0.412	
243	0.415	0.413	0.417
852	0.409	0.414	0.420
548	0.418	0.418	0.424
1158	0.420	0.413	0.417
1463	0.416	0.411	

**Table 22: Results for Cd by ID-ICP-MS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
18	1.615	1.627	
77	1.620	1.645	
156	1.623	1.628	
164	1.615	1.618	
187	1.627	1.633	
243	1.620	1.615	1.625
852	1.610	1.609	1.616
548	1.633	1.617	1.626
1158	1.614	1.618	1.617
1463	1.616	1.621	1.617



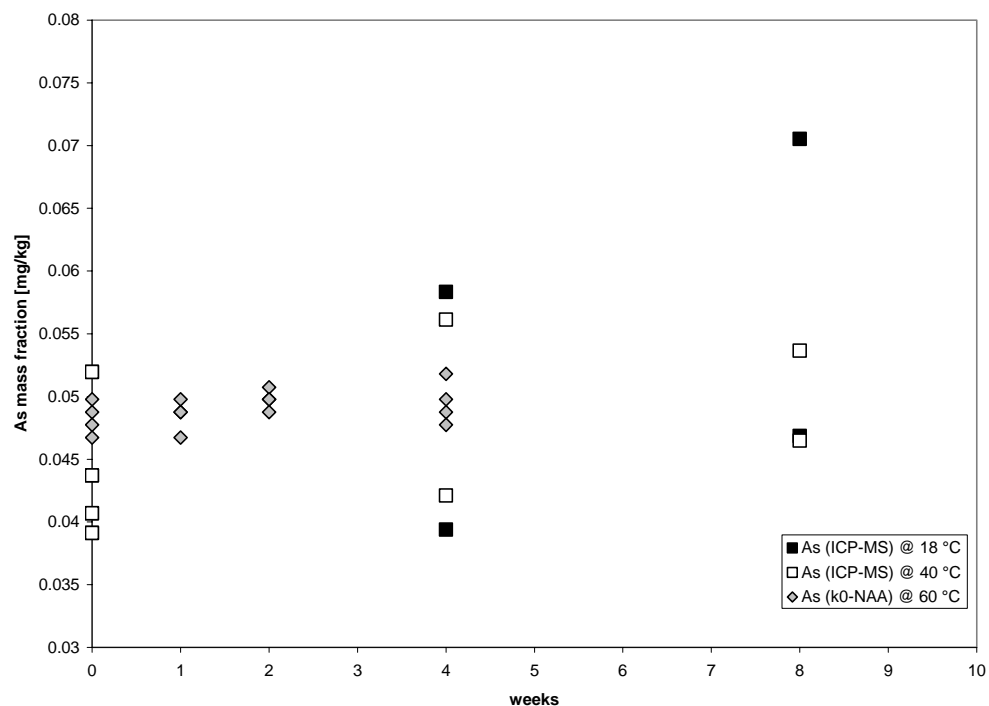
**Table 23: Results for Cu by ID-ICP-MS (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2	Replicate 3
18	2.79	2.80	
77	3.06	2.81	
156	2.79	2.84	
164	2.80	2.81	
187	2.84	2.83	
243	2.81	2.83	2.84
852	2.78	2.81	2.80
548	2.83	2.83	2.87
1158	2.83	2.80	2.81
1463	2.81	2.79	2.80

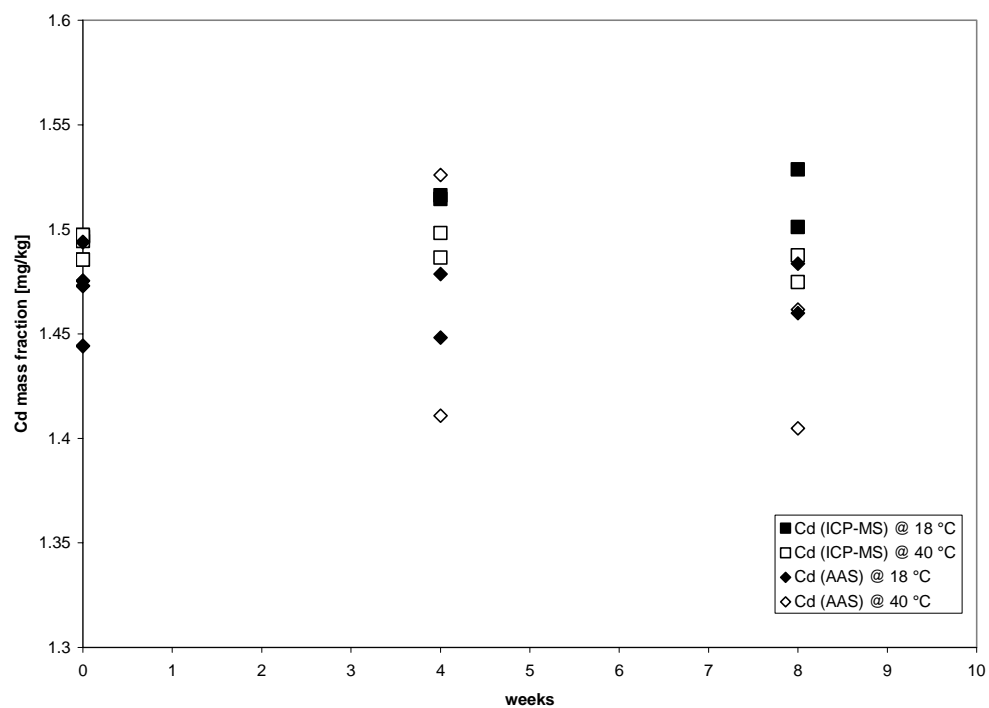
**Table 24: Results for As by k<sub>0</sub>-NAA (results are expressed in mg/kg)**

Bottle no.	Replicate 1	Replicate 2
140	0.049	0.049
158	0.048	0.047
182	0.049	0.047
331	0.052	0.047
372	0.050	0.049
425	0.048	0.052
520	0.047	0.046
567	0.048	0.049
615	0.048	0.048
710	0.049	0.049
782	0.048	0.046
805	0.050	0.046
832	0.050	0.049
900	0.049	0.048
979	0.050	0.049
1156	0.051	0.050
1350	0.052	0.050
1445	0.048	0.049

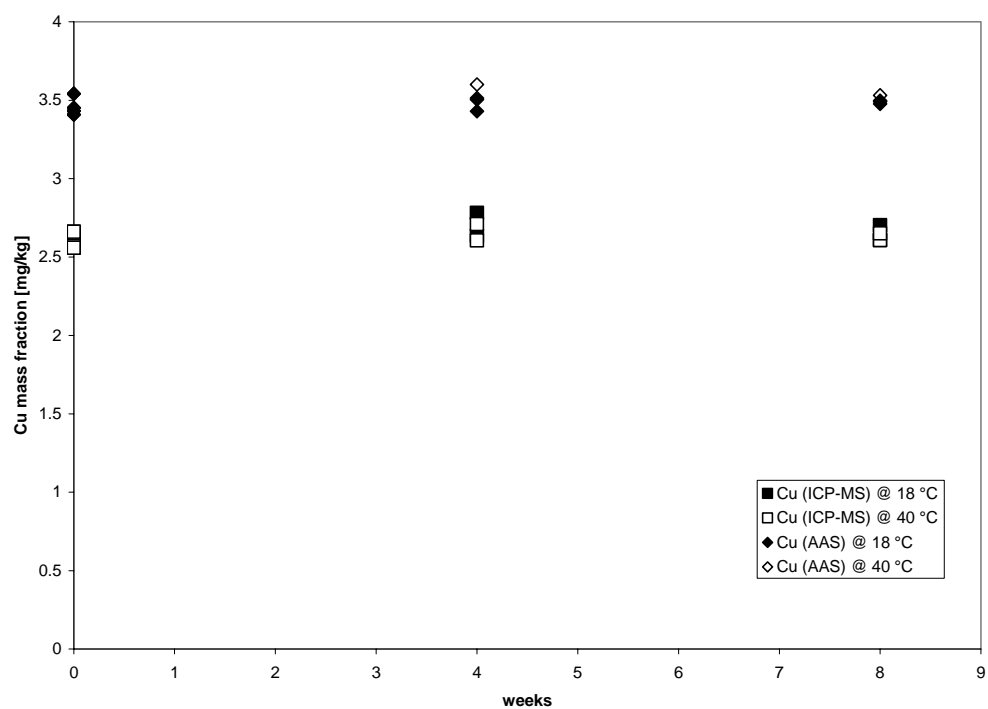
### Annex 3: Data from stability studies



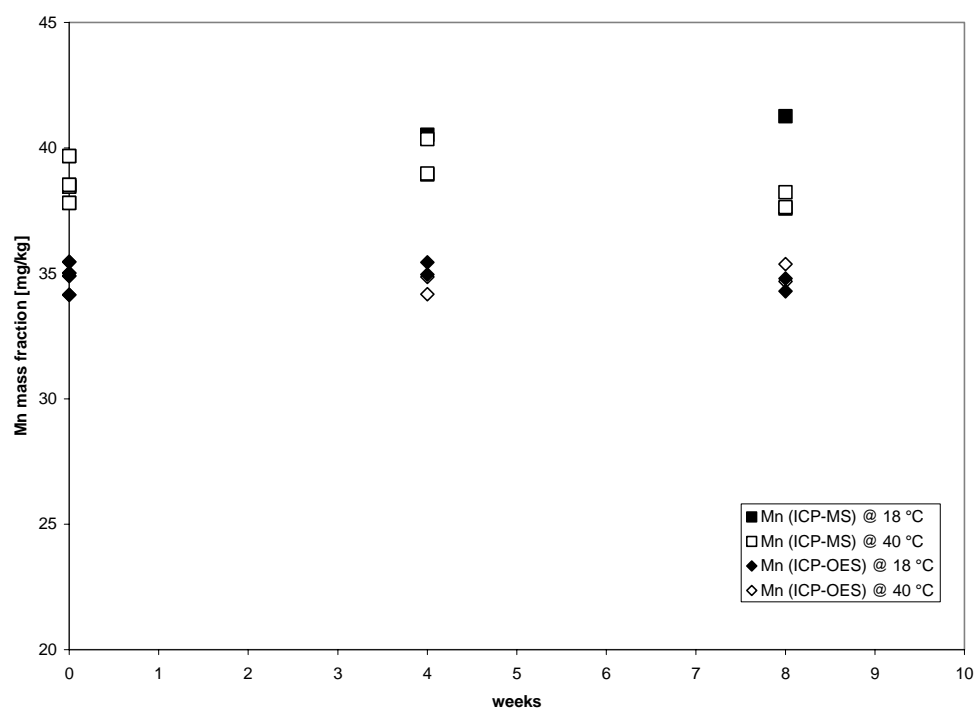
**Figure 2: Short-term stability graph for As**



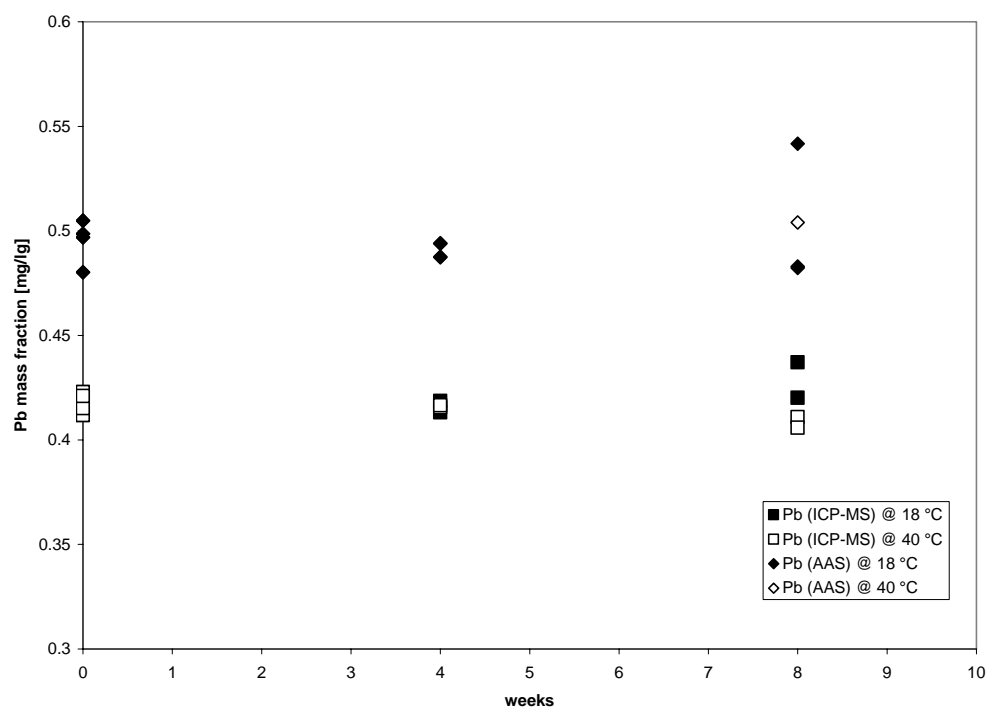
**Figure 3: Short-term stability graph for Cd**



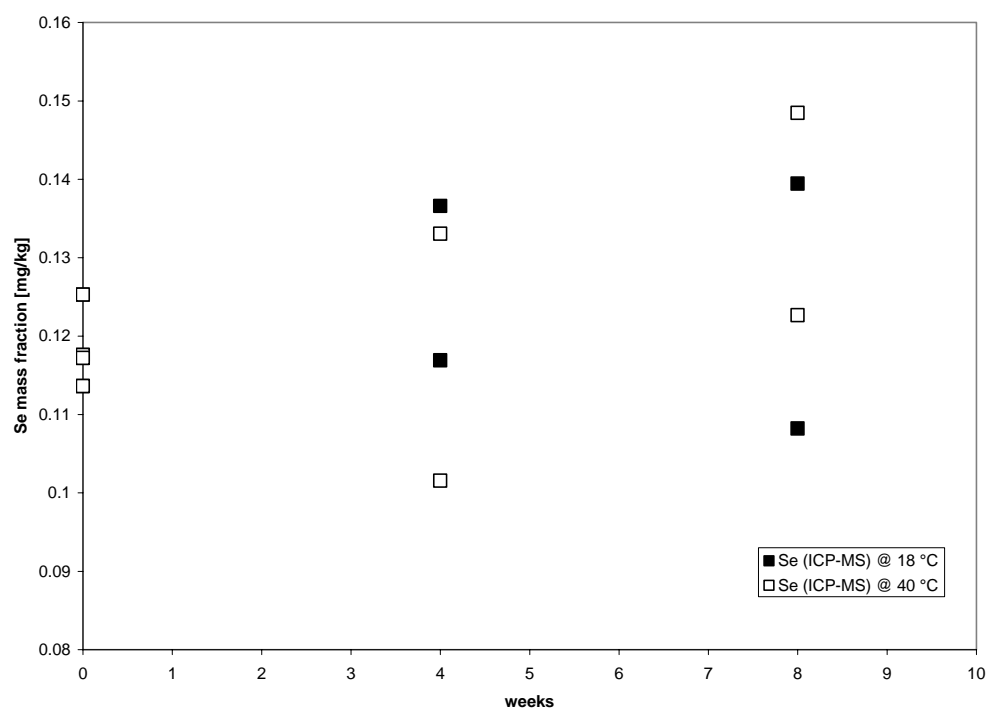
**Figure 4: Short-term stability graph for Cu**



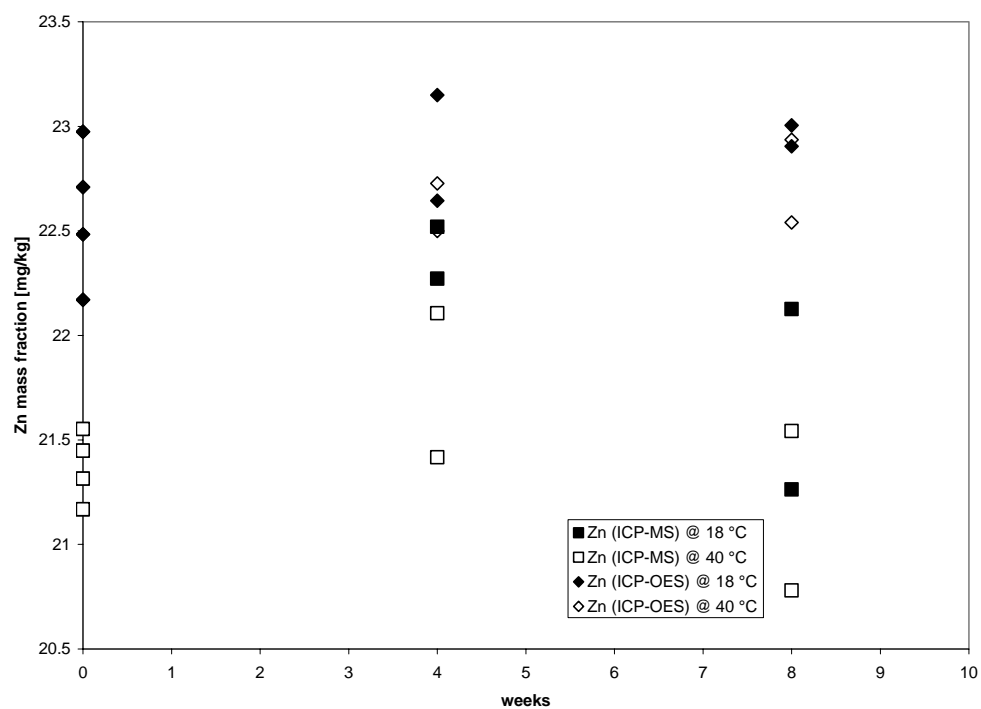
**Figure 5: Short-term stability graph for Mn**



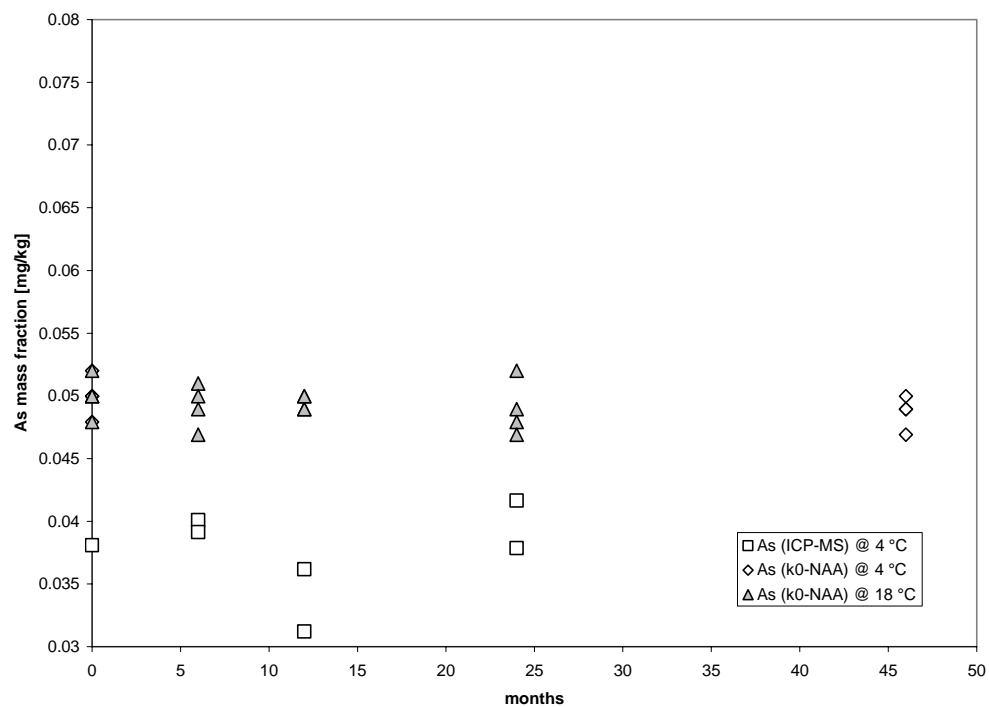
**Figure 6: Short-term stability graph for Pb**



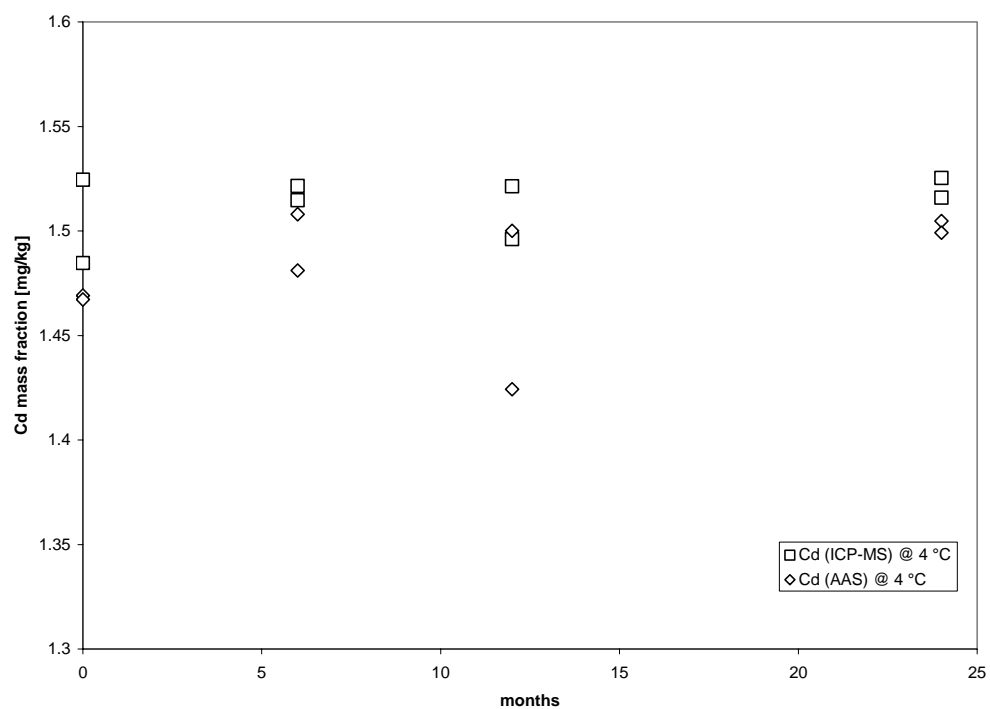
**Figure 7: Short-term stability graph for Se**



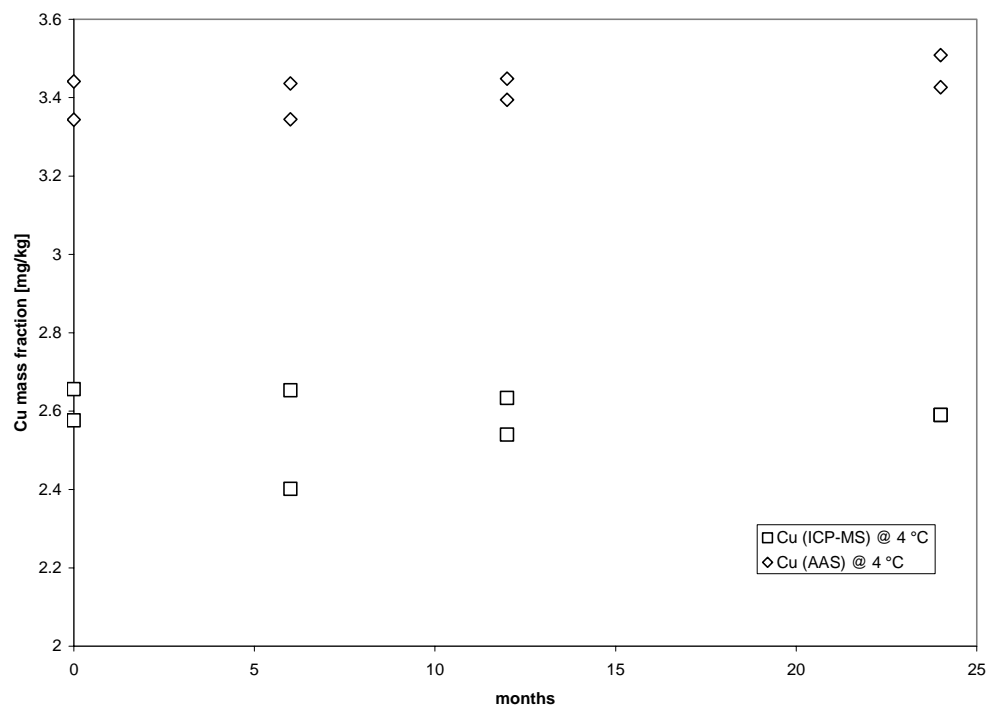
**Figure 8: Short-term stability graph for Zn**



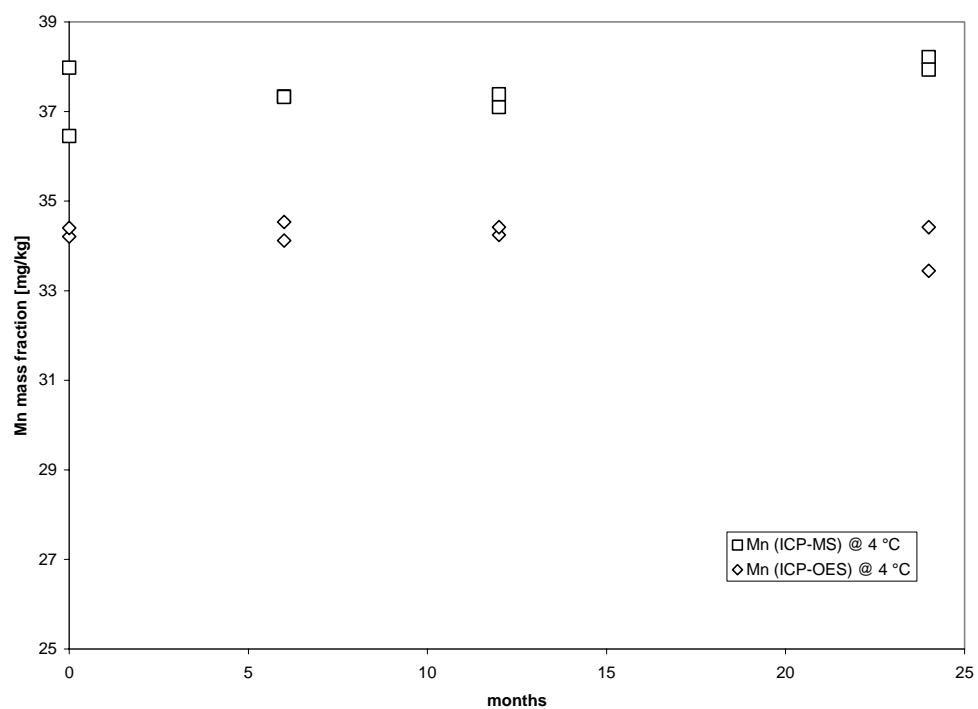
**Figure 9: Long-term stability graph for As**



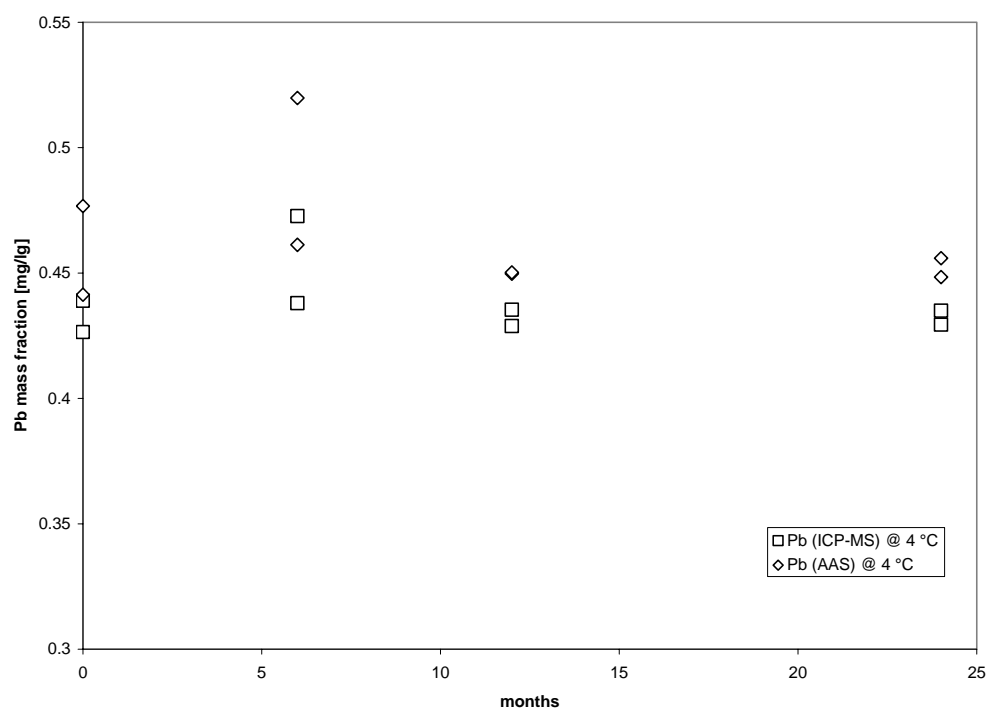
**Figure 10: Long-term stability graph for Cd**



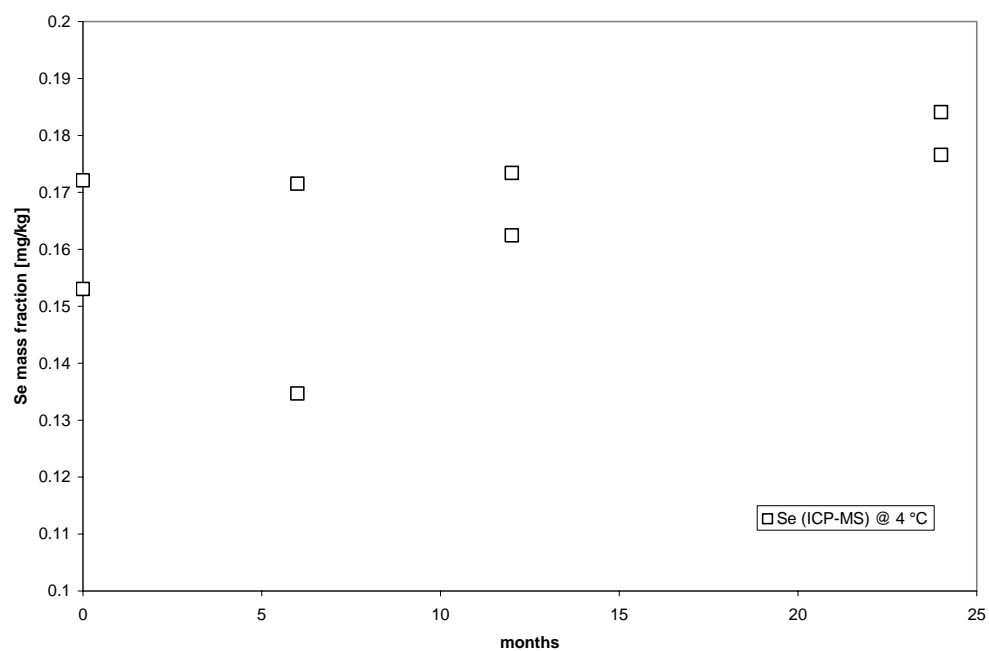
**Figure 11: Long-term stability graph for Cu**



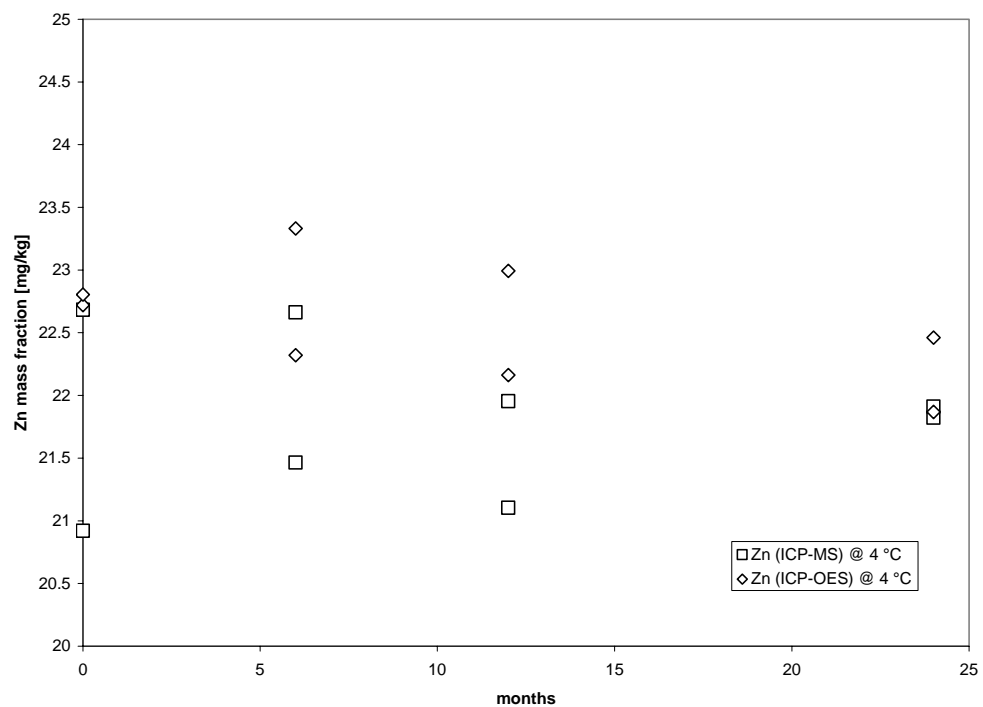
**Figure 12: Long-term stability graph for Cu**



**Figure 13: Long-term stability graph for Pb**



**Figure 14: Long-term stability graph for Se**



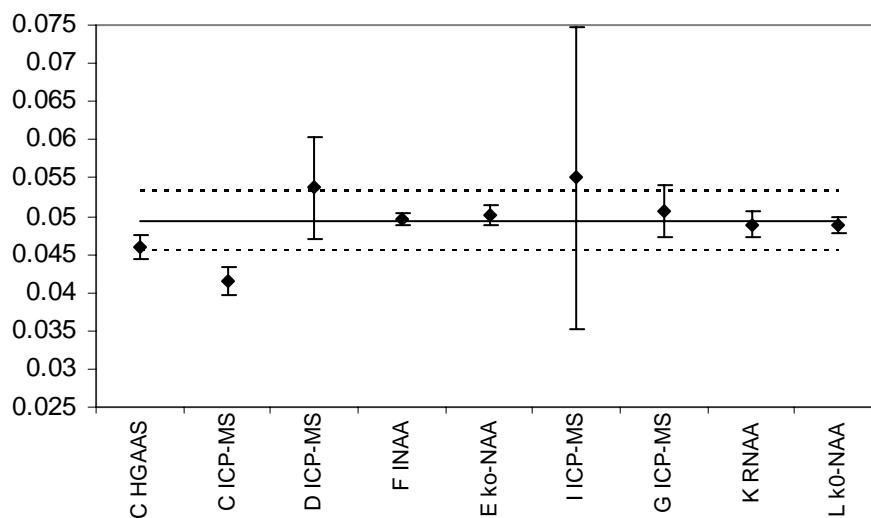
**Figure 15: Long-term stability graph for Zn**



#### Annex 4: Data from characterisation studies

**Table 25: Individual results as reported for As (data expressed in mg/kg). Data not included in the final evaluation is indicated by a grey background. Results for Lab C are presented as individual results, but are only used as one set of data for the evaluation. sd: standard deviation**

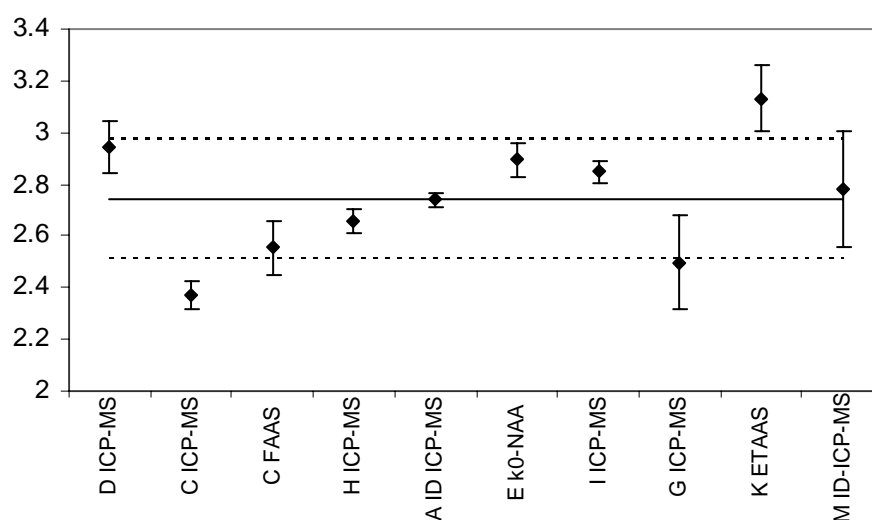
Labcode	Results of the replicate measurements (mg/kg)						mean	sd
C HGAAS	0.047	0.044	0.047	0.047	0.044	0.047	0.046	0.0015
C ICP-MS	0.044	0.041	0.043	0.04	0.039	0.042	0.0415	0.0019
D ICP-MS	0.067	0.05	0.05	0.05	0.053	0.052	0.0537	0.0067
H ICP-MS	0.0744	0.0336	0.0487	0.0579	0.0605	0.0368		
I ICP-MS	0.03	0.07	0.03	0.06	0.07	0.07	0.055	0.0197
G ICP-MS	0.05	0.053	0.045	0.054	0.053	0.049	0.0507	0.0034
J INAA	0.0746	0.0684	0.0725	0.0780	0.0746	0.0734		
F INAA	0.0503	0.0487	0.0488	0.0489	0.0503	0.0503	0.0495	0.0008
K RNAA	0.0496	0.0492	0.051	0.046	0.0483	0.0491	0.0489	0.0017
E k <sub>0</sub> -NAA	0.0480	0.0498	0.0507	0.0503	0.0497	0.0523	0.0501	0.0014
L k <sub>0</sub> -NAA	0.0492	0.0488	0.0475	0.0500	0.0480	0.0498	0.0489	0.0010



**Figure 16: Results for As as used for characterisation. Uncertainty bars represent the 95 % confidence interval of each lab's results. x-axis expressed in mg/kg ( — : certified value, - - : expanded uncertainty of the certified value)**

**Table 26: Individual results as reported for Cu (data expressed in mg/kg). Data not included in the final evaluation is indicated by a grey background. Results for Lab C are presented as individual results, but are only used as one set of data for the evaluation. sd: standard deviation**

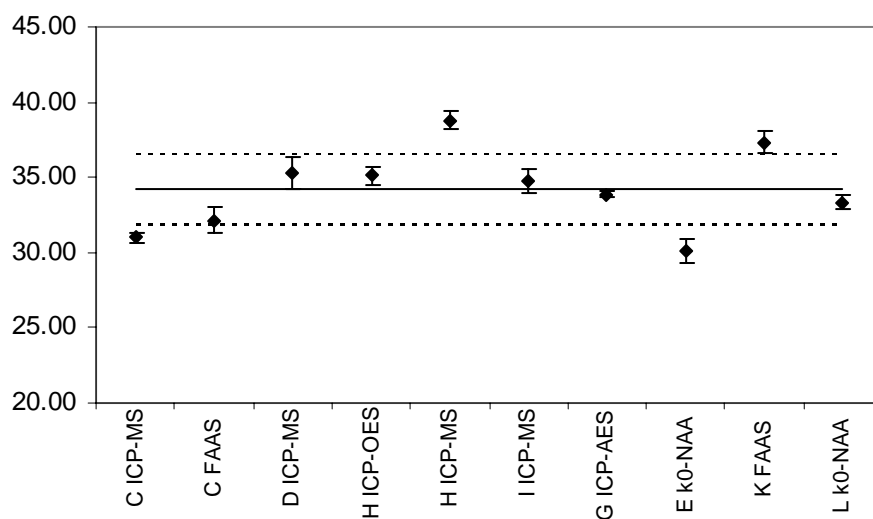
Labcode	Results of the replicate measurements (mg/kg)						mean	sd
D ICP-MS	3.00	3.11	2.93	2.86	2.90	2.84	2.94	0.101
C ICP-MS	2.41	2.40	2.38	2.30	2.43	2.30	2.37	0.057
H ICP-MS	2.653	2.703	2.654	2.574	2.645	2.706	2.656	0.048
I ICP-MS	2.90	2.86	2.84	2.81	2.79	2.89	2.848	0.044
G ICP-MS	2.34	2.40	2.38	2.65	2.43	2.79	2.498	0.180
A ID ICP-MS	2.724	2.716	2.721	2.765	2.783	2.725	2.739	0.028
M ID-ICP-MS	2.792	2.799	3.082	2.809	2.188	2.835		
	2.801	2.807	2.843	2.83			2.779	0.225
C FAAS	2.60	2.39	2.58	2.59	2.69	2.48	2.555	0.105
H GFAAS	3.544	3.766	3.799	3.579	3.472	3.759		
K ETAAS	3.27	3.25	3.00	3.22	2.99	3.06	3.132	0.129
J INAA	6.34	6.56	6.01	6.4	6.64	6.03		
E k <sub>0</sub> -NAA	2.914	2.861	2.944	2.829	2.995	2.826	2.895	0.068



**Figure 17: Results for Cu as used for characterisation. Uncertainty bars represent the 95 % confidence interval of each lab's results. x-axis expressed in mg/kg ( — : certified value, - - : expanded uncertainty of the certified value)**

**Table 27: Individual results as reported for Mn (data expressed in mg/kg). Results for Lab C are presented as individual results, but are only used as one set of data for the evaluation. sd: standard deviation**

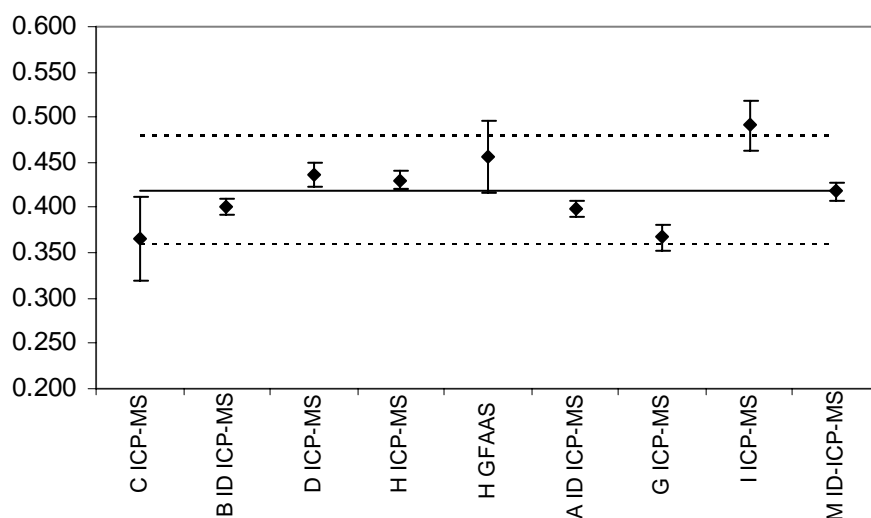
Labcode	Results of the replicate measurements (mg/kg)						mean	sd
C FAAS	32.7	31.9	30.6	33.1	32.1	32.3	32.12	0.86
K FAAS	37.3	37.6	36.9	36.4	37.3	38.5	37.33	0.71
C ICP-MS	31.0	31.5	30.6	30.9	30.8	31.1	30.98	0.31
D ICP-MS	35.4	35.1	36.9	33.8	34.8	35.9	35.32	1.05
H ICP-MS	38.37	38.34	38.68	38.60	39.03	39.82	38.81	0.56
I ICP-MS	35.4	34.6	34.9	34.5	33.5	35.6	34.75	0.75
H ICP-OES	35.66	34.04	35.13	35.70	35.43	34.64	35.10	0.65
G ICP-AES	33.9	34.1	33.7	34	33.9	33.7	33.88	0.16
J INAA	34.71	35.74	35.54	34.09	33.89	35.12		
E k <sub>0</sub> -NAA	28.65	29.61	30.33	30.46	30.52	31.03	30.10	0.84
L k <sub>0</sub> -NAA	34.16	33.64	32.97	33.06	33.22	32.95	33.33	0.47



**Figure 18: Results for Mn as used for characterisation. Uncertainty bars represent the 95 % confidence interval of each lab's results. x-axis expressed in mg/kg ( — : certified value, - - : expanded uncertainty of the certified value)**

**Table 28: Individual results as reported for Pb (data expressed in mg/kg). Data not included in the final evaluation is indicated by a grey background. Results for Lab C are presented as individual results, but are only used as one set of data for the evaluation. sd: standard deviation**

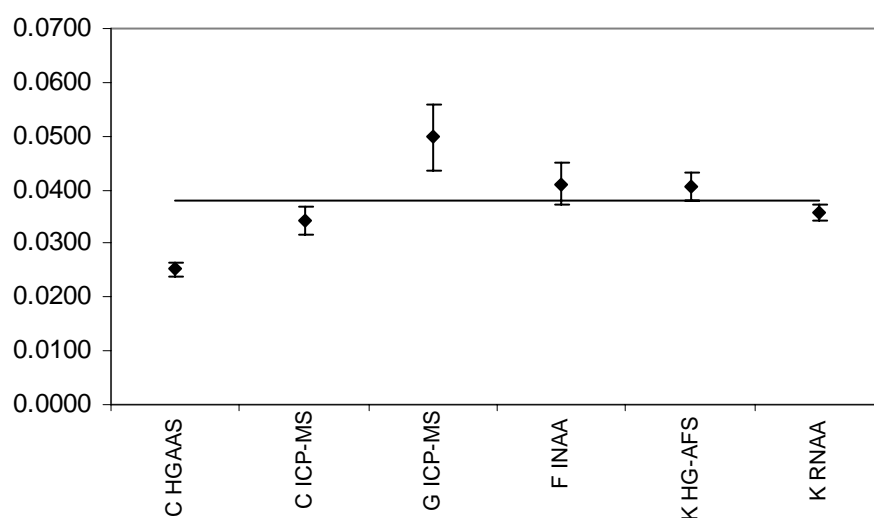
Labcode	Results of the replicate measurements (mg/kg)						mean	sd
C FAAS	<=1.0	<=1.0	<=1.0	<=1.0	<=1.0	<=1.0		
H GFAAS	0.5341	0.4332	0.4418	0.4593	0.4465	0.4217	0.456	0.040
C ICP-MS	0.412	0.372	0.32	0.398	0.391	0.297	0.365	0.046
D ICP-MS	0.44	0.46	0.42	0.43	0.43	0.44	0.437	0.014
H ICP-MS	0.4441	0.4245	0.4274	0.4161	0.4403	0.4331	0.431	0.010
G ICP-MS	0.372	0.386	0.355	0.370	0.372	0.348	0.367	0.014
I ICP-MS	0.469	0.483	0.500	0.518	0.523	0.454	0.491	0.027
A ID ICP-MS	0.3891	0.4054	0.4084	0.3923	0.3952	0.4038	0.399	0.008
B ID ICP-MS	0.4185	0.3984	0.3999	0.4056	0.3935	0.3941	0.402	0.009
M ID-ICP-MS	0.413	0.415	0.435	0.424	0.419	0.418		
	0.403	0.411	0.427	0.412			0.418	0.009



**Figure 19: Results for Pb as used for characterisation. Uncertainty bars represent the 95 % confidence interval of each lab's results. x-axis expressed in mg/kg ( — : certified value, - - : expanded uncertainty of the certified value)**

**Table 29: Individual results as reported for Se (data expressed in mg/kg). Data not included in the final evaluation is indicated by a grey background. Results for Lab C are presented as individual results, but are only used as one set of data for the evaluation. sd: standard deviation**

Labcode	Results of the replicate measurements (mg/kg)						mean	sd
C HGAAS	0.027	0.025	0.024	0.026	0.025	0.024	0.0252	0.0012
K HG-AFS	0.042	0.042	0.040	0.043	0.041	0.036	0.0407	0.0025
C ICP-MS	0.039	0.033	0.032	0.035	0.035	0.032	0.0343	0.0027
D ICP-MS	<0,1	<0,1	<0,1	<0,1	<0,1	<0,1		
H ICP-MS	0.1614	0.1437	0.1910	0.1527	0.1301	0.1842		
I ICP-MS	<0.1	<0.1	<0.1	<0.1	<0.1	<0.1		
G ICP-MS	0.057	0.044	0.054	0.052	0.041	0.051	0.0498	0.0061
J INAA	0.4275	0.4470	0.4460	0.4810	0.4429	0.4378		
F INAA	0.0434	0.0399	0.0386	0.036	0.0412	0.0472	0.0411	0.0039
K RNAA	0.0336	0.0353	0.0369	0.038	0.0348	0.0362	0.0358	0.0016



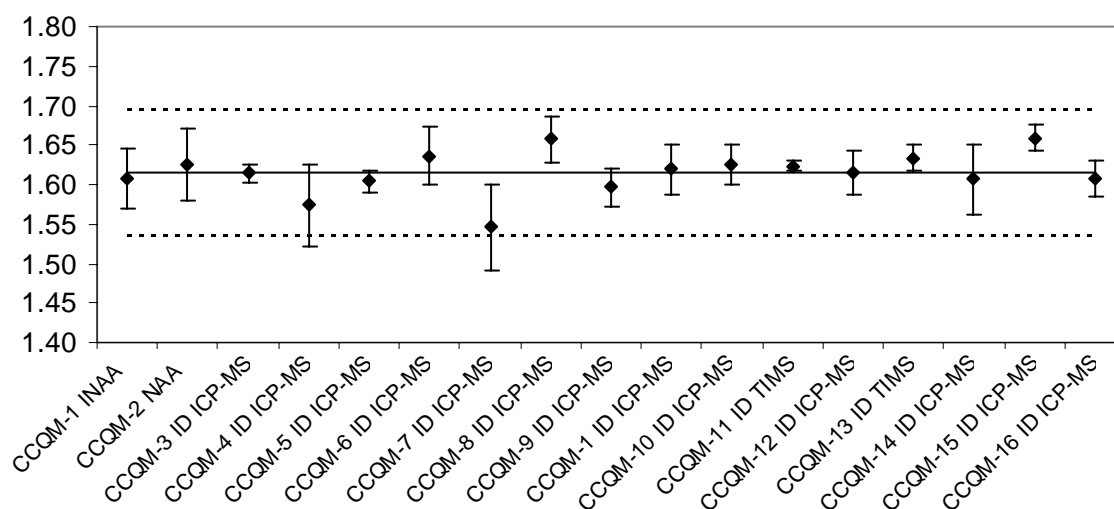
**Figure 20: Results for Se as used for characterisation. Uncertainty bars represent the 95 % confidence interval of each lab's results. x-axis expressed in mg/kg ( — : "additional material information" value)**

**Table 30: Individual results as reported for Cd (data expressed in mg/kg).**

Labcode	Mean <sup>1)</sup>	Expanded uncertainty as reported by laboratory <sup>2)</sup>
CCQM-1 INAA	1.61	0.04
CCQM-2 NAA	1.63	0.04
CCQM-3 ID ICP-MS	1.614	0.011
CCQM-4 ID ICP-MS	1.57	0.05
CCQM-5 ID ICP-MS	1.604	0.015
CCQM-6 ID ICP-MS	1.64	0.04
CCQM-7 ID ICP-MS	1.55	0.06
CCQM-8 ID ICP-MS	1.66	0.03
CCQM-9 ID ICP-MS	1.60	0.02
CCQM-1 ID ICP-MS	1.62	0.03
CCQM-10 ID ICP-MS	1.63	0.02
CCQM-12 ID ICP-MS	1.62	0.03
CCQM-14 ID ICP-MS	1.61	0.04
CCQM-15 ID ICP-MS	1.659	0.017
CCQM-16 ID ICP-MS	1.61	0.02
CCQM-11 ID TIMS	1.623	0.006
CCQM-13 ID TIMS	1.633	0.017

<sup>1)</sup>In the CCQM study, laboratories were free to choose the number of replicates, and they were not asked to report results for their individual replicate measurements. Only the resulting laboratory mean was reported.

<sup>2)</sup>Results and uncertainties were all reported in nmol/g, only some laboratories also reported results in mg/kg. Results were converted in mg/kg during this evaluation.



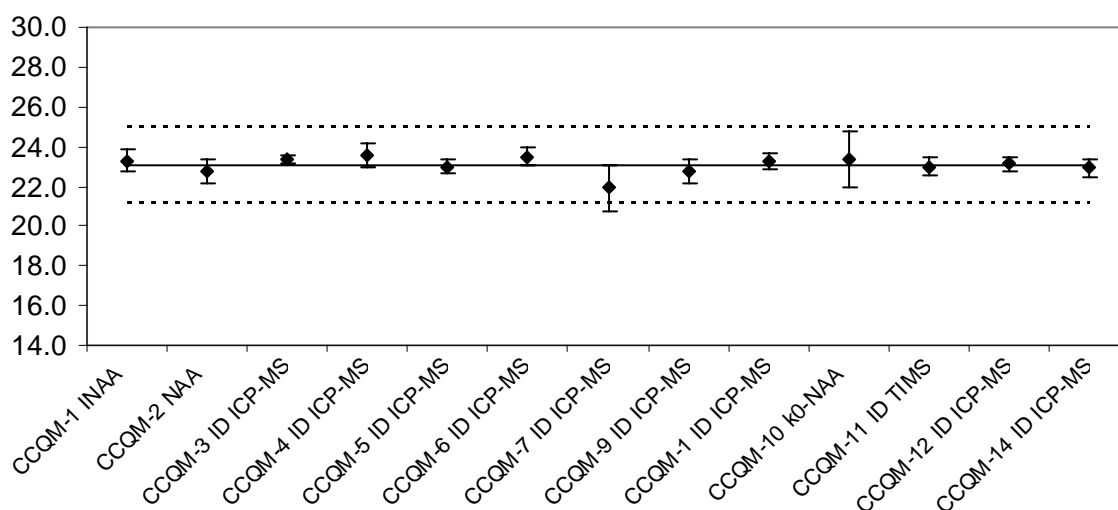
**Figure 21: Results for Cd as used for characterisation. Uncertainty bars represent the the expanded uncertainties of the measurements as reported by the participating laboratories. y-axis expressed in mg/kg ( — : certified value, - - : expanded uncertainty of the certified value)**

**Table 31: Individual results as reported for Zn (data expressed in mg/kg).**

Labcode	Mean <sup>1)</sup>	Expanded uncertainty as reported by laboratory <sup>2)</sup>
CCQM-1 INAA	23.3	0.6
CCQM-2 NAA	22.8	0.6
CCQM-10 k0-NAA	23.4	1.4
CCQM-3 ID ICP-MS	23.4	0.2
CCQM-4 ID ICP-MS	23.5	0.6
CCQM-5 ID ICP-MS	23.0	0.4
CCQM-6 ID ICP-MS	23.5	0.4
CCQM-7 ID ICP-MS	21.9	1.2
CCQM-9 ID ICP-MS	22.8	0.6
CCQM-1 ID ICP-MS	23.2	0.4
CCQM-12 ID ICP-MS	23.1	0.4
CCQM-14 ID ICP-MS	22.9	0.5
CCQM-11 ID TIMS	23.0	0.4

<sup>1)</sup>In the CCQM study, laboratories were free to choose the number of replicates, and they were not asked to report results for their individual replicate measurements. Only the resulting laboratory mean was reported.

<sup>2)</sup>Results and uncertainties were all reported in nmol/g, only some laboratories also reported results in mg/kg. Results were converted in mg/kg during this evaluation.



**Figure 22: Results for Zn as used for characterisation. Uncertainty bars represent the the expanded uncertainties of the measurements as reported by the participating laboratories. y-axis expressed in mg/kg ( — : certified value, - - : expanded uncertainty of the certified value)**

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The certification of the Mass Fractions of As, Cd, Cu, Mn, Pb, Se and Zn in rice flour, IRMM-804

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**Abstract**

This report describes the preparation and certification of a rice flour Certified Reference Material (CRM) IRMM-804. The CRM was processed and certified by the European Commission, Directorate General Joint Research Centre, Institute for Reference Materials and Measurements (IRMM), Geel, Belgium.

The CRM was prepared from rice grown on Cd enriched water. After milling the resulting powder was filled in glass bottles containing 15 g of sample.

Certification of the CRM included testing of the homogeneity and stability of the material as well as the characterisation using an intercomparison approach.

The new CRM has been certified for its content of As, Cd, Cu, Mn, Pb and Zn, while the Se mass fraction is only given as "additional material information".

The main purpose of the material is to assess method performance, i.e. for checking accuracy of analytical results. As any reference material, it can also be used for control charts or validation studies.

	Mass Fraction	
	Certified value <sup>1)</sup> [mg/kg]	Uncertainty <sup>2)</sup> [mg/kg]
As	0.049	0.004
Cu	2.74	0.24
Mn	34.2	2.3
Pb	0.42	0.07
Cd	1.61	0.07
Zn	23.1	1.9
1) Unweighted mean value of the means of accepted sets of data, each set being obtained in a different laboratory and/or with a different method of determination. The certified values are traceable to the SI.		
2) Expanded uncertainty with a coverage factor k = 2 according to the Guide for the Expression of Uncertainty in Measurement, corresponding to a level of confidence of about 95 %.		





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